Phenotypic performance of four stevia genotypes in the Alto Vale do Itajaí region, Brazil

Rendimiento fenotípico de cuatro genotipos de estevia en la Región del Alto Vale do Itajaí, Brasil

ABSTRACT

An evaluation of four stevia genotypes for biomass yield, stevioside and rebaudioside A content and yield under decreasing photoperiod conditions was carried out in the Alto Vale do Itajaí region, located in the State of Santa Catarina (SC), Brazil. This field experiment was conducted at Site São Miguel, a farm located in the city of Lontras (SC), Brazil, under conditions of decreasing photoperiods, with a variation of 13.72 h of light at experiment implantation to 12.57 h of light at the end of the evaluations. The treatments consisted of four genotypes (G4, G8, G9 and G12) provided by EMBRAPA-CENARGEN. A randomized complete block design with four treatments (stevia genotypes) and four replications was used. Each plot consisted of 21 plants, and the floor area had five plants. G12 had the highest leaf dry weight (LDW), total leaf area, leaf area index, leaf area ratio and specific leaf area of all the genotypes. G4 and G12 were equal for LDW and were higher than the other genotypes, with yields of 755.6 and 836.4 kg ha⁻¹, respectively. The stevioside content was highest in G12 (200.07 mg g⁻¹). G8 and G9 were similar for rebaudioside A content (64.77 and 49.05 mg g⁻¹, respectively). The rebaudioside A: stevioside ratio was highest in G8 (0.44 g g⁻¹). No genotype had a rebaudioside A: stevioside ratio suitable for industry requirements.

Additional key words: photoperiodicity; physiological response; genotypes; sweeteners; Stevia rebaudiana (Bert.) Bertoni; stevioside; rebaudioside.

1 Universidade Federal do Paraná, Doutorando do programa de Pós Graduação em Produção Vegetal, Curitiba; Instituto Federal Catarinense, campus Rio do Sul, Rio do Sul (Brazil). ORCID Debarba, R.J.: 0000-0002-0074-8866
2 Universidade Federal do Paraná, Departamento de Fitotecnia e Fitossanitarismo, Curitiba (Brazil). ORCID Deschamps, C.: 0000-0003-0786-0532
3 Universidade do Sul de Santa Catarina, Grupo de Pesquisa em Tecnologia Farmacêutica, Tubarão (Brazil). ORCID Kanis, L.A.: 0000-0001-7600-7550; ORCID Moterle, D.: 0000-0002-9383-4295
5 Corresponding author. romulo.debarba@ifc.edu.br
The demand for sweeteners for dietary and pharmaceutical purposes is growing. Stevia (*Stevia rebaudiana* Bert.) contributes to the supply of natural and non-carcinogenic sweeteners (Anton *et al*., 2010). Stevia (Asteraceae), native to Brazil and Paraguay, has been used for its medicinal and dietary properties, derived from its glycosides stevioside and rebaudioside A, both of which are 300 times sweeter than sucrose (Espita *et al*., 2009).

The relationship between stevioside and rebaudioside A is an important variable for analysis in new stevia genotypes. According to Mota *et al.* (2015), the industry prefers genotypes that have a ratio close to 1 g g\(^{-1}\) because stevioside has a low water solubility and bitter residual taste, while rebaudioside A is more soluble and has no residual taste.

Stevioside and rebaudioside A are more concentrated in leaves and are diterpene glycosides that are synthesized in the same mevalonate pathway as gibberellic acid (Jarma *et al*., 2010).

The cultivation of stevia in Brazil is incipient and does not meet the growing domestic demand. Brazil imported more than US$8 million and exported US$2.7 million in stevia in 2013 (MDIC, 2019).

Stevia has potential for cultivation in the Alto Vale do Itajaí region, Santa Catarina. The increasing demand for glycosides warrants studies for the selection of new genotypes with productive potential, local adaptation, and stevioside and rebaudioside A ratios required by the industry (Mota *et al*., 2015).

In southern Brazil, studies by the Universidade Federal do Paraná (UFPR) with unevaled genotypes provided by EMBRAPA-CENARGEN collected in different regions of Brazil, showed that stevia can be grown in the Curitiba region, State of Paraná (Francisco, 2015), justifying exploratory research on the genotypes provided by EMBRAPA-CENARGEN in areas with different soils and climates, such as those in the Alto Vale do Itajaí region (SC).

Because of the lack of agronomic information and genotypes adapted to the Alto Vale do Itajaí region (SC), this study aimed to evaluate four different stevia genotypes provided by EMBRAPA-CENARGEN...
in terms of dry weight yield, glycoside content and yield, and rebaudioside A: stevioside ratio.

**MATERIAL AND METHODS**

This study was carried out at Sítio São Miguel, located in the city of Lontras (SC), Brazil (27º 9’ 58” S, 49º 32’ 31” W, altitude of 475 m). The climate was Cfa according to the Köppen classification; climate data during the experiment were determined (Tab. 1).

The soil in the experiment area was classified as Argissolo Amarelo distrófico típico (EMBRAPA, 2006). Soil analysis data were collected (Tab. 2).

The soil was corrected according to laboratory analysis results. The corrections were based on the Fertilization and Liming Manual for the states of Santa Catarina and Rio Grande do Sul (SBCS and CQFS, 2016). The soil was corrected with phosphorus and potassium fertilization when preparing the beds, 15 d prior to transplanting the seedlings. A rotary tiller was used to prepare the soil of the experiment area.

The fertilization was carried out with doses (kg ha⁻¹): 90 N, 68 P₂O₅ and 110 K₂O, supplied by urea, superphosphate and potassium chloride, respectively. A split application of urea was done at transplanting and 30 d after transplanting, according to Lima Fiho (2004). The genotypes used in this study were supplied by EMBRAPA - CENARGEN (Brasília, DF). The genotypes have distinct morphological and productive characteristics, as evidenced by previous studies at UFPR (Francisco, 2015).

The seedlings were obtained from branches of mother plants located at Fazenda Canguiri - UFPR – Pinhais (PR), Brazil, based on the methodology of Carvalho and Zaidan (1995) and using IBA (Indol butyric acid) at a concentration of 2000 mg L⁻¹. The seedlings were produced in commercial substrate (Macrofétil®) on the premises of the Instituto Federal Catarinense (IFC) - Rio do Sul (SC) campus. At transplanting, the seedlings were subjected to apical bud breaking, according to EMBRAPA (2004), which consisted of cutting the seedlings at a height of 5 cm from the soil.

The seedlings were transplanted in beds prepared with a raised bed planter. The experiment was installed on December 12, 2014. The spacing used was 0.25 x 0.50 m - 80,000 plants/ha (SBCS and CQFS, 2016).

The treatments consisted of Genotype 4 (G4), Genotype 8 (G8), Genotype 9 (G9) and Genotype 12 (G12). A randomized complete block design with four treatments (stevia genotypes) and four replicates was used. Each plot consisted of 21 plants, with 5 plants per useful area. Plants were harvested when they were at 5% flowering (Lima Fiho, 2004), which occurred at different times for each genotype.

The following characteristics were evaluated: leaf dry weight (LDW), stem dry weight (SDW), branch

### Table 1. Temperature, humidity and photoperiod during the experiment. Rio do Sul (2014/2015).

<table>
<thead>
<tr>
<th>Climate data</th>
<th>December</th>
<th>January</th>
<th>February</th>
<th>March</th>
</tr>
</thead>
<tbody>
<tr>
<td>Maximum temperature (°C)</td>
<td>30.7</td>
<td>30.9</td>
<td>30.6</td>
<td>26.4</td>
</tr>
<tr>
<td>Minimum temperature (°C)</td>
<td>13.9</td>
<td>15.9</td>
<td>15.6</td>
<td>12.2</td>
</tr>
<tr>
<td>Average temperature (°C)</td>
<td>20.5</td>
<td>20.5</td>
<td>20.9</td>
<td>18.3</td>
</tr>
<tr>
<td>Relative air humidity (%)</td>
<td>89</td>
<td>91</td>
<td>93</td>
<td>94</td>
</tr>
<tr>
<td>Photoperiod (h)</td>
<td>13.72</td>
<td>13.68</td>
<td>13.24</td>
<td>12.57</td>
</tr>
</tbody>
</table>


### Table 2. Soil chemical characteristics of the experiment area.

<table>
<thead>
<tr>
<th>pH SMP</th>
<th>Al⁻³</th>
<th>H⁺ + Al⁺³</th>
<th>Ca⁺²</th>
<th>Mg⁺²</th>
<th>CEC</th>
<th>K⁺</th>
<th>P</th>
<th>OM</th>
<th>V</th>
</tr>
</thead>
<tbody>
<tr>
<td>cmol dm⁻³</td>
<td>mg dm⁻³</td>
<td>%</td>
<td>%</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>6.0</td>
<td>0.3</td>
<td>4.5</td>
<td>2.9</td>
<td>2.0</td>
<td>9.42</td>
<td>32.0</td>
<td>10.8</td>
<td>1.3</td>
<td>52.19</td>
</tr>
</tbody>
</table>
dry weight (BDW), total dry weight (TDW), main branch length, number of secondary branches, number of tertiary branches, leaf area (LA), leaf area index (LAI), specific leaf area (SLA), leaf area ratio (LAR), leaf weight ratio (LWR), stevioside and rebaudioside A yield, stevioside and rebaudioside A contents, and rebaudioside A: stevioside- ratio. The plants of the useful plot were cut 5 cm from the ground (Lima Fiho, 2004) and taken to the Plant Physiology Laboratory of IFC (Rio do Sul campus), where the leaves were separated from the stems. The LDW and SDW were determined on a digital scale after drying the leaves and stems at 50 ºC until constant weight was reached (Espita et al., 2009). A tape measure was used to determine the height of the main branch. The number of secondary and tertiary branches was counted.

The leaf area (LA), leaf area index (LAI), specific leaf area (SLA), leaf area ratio (LAR) and leaf weight ratio (LWR) were determined using the methodologies described by Cunha et al. (2010) using an artisanal leaf disc cutter with an area of 10 mm, precision electronic scale (Gehaka ® AG 220S) and air circulation oven (ACB Labor®).

To quantify the stevioside and rebaudioside A yield, samples of 1 g of leaf tissue were collected from plants in the useful area of each treatment. A leaf tissue sample was randomly collected from the LDW of the plants that made up the useful area of the plot. We used the extraction and quantification methodology described by Kolb et al. (2001), with modifications. The modification consisted of using 0.3 mL of the sample extracted from the leaves of each treatment, adding 0.7 mL of HPLC grade acetonitrile. Dry leaf tissue samples were placed in 250 mL Erlenmeyer flasks with 100 mL of 70% ethanol. The solution was heated to 70 ºC and stirred for 30 min. After cooling, a 10 mL aliquot was filtered (quantitative filter paper and a 0.22 µm nylon syringe filter). We used 0.3 mL of the sample extracted from the leaves of each treatment by adding 0.7 mL of HPLC grade acetonitrile. From this dilution, 20 µL were injected for further analysis in a High Performance Liquid Chromatograph (Shimadzu CBM-10A) containing a Phenomenex Luna® 5 µm NH2 100 Å, 250 x 4.6 mm column. Elution was at room temperature in isocratic mode using a mixture of acetonitrile-distilled water (80:20, v/v) as a solvent and a flow rate of 2 mL min⁻¹. Detection was done by UV at 210 nm with sensitivity adjusted to 0.04 AUFS. Readings were taken in triplicate. The quantification of each metabolite was obtained by converting the area of the curve corresponding to retention time using a previously established calibration curve. The standard solution for obtaining the calibration curve was 1.0 g L⁻¹ stevioside and rebaudioside A in methanol (Kolb et al., 2001). The conversion was expressed in mg g⁻¹.

The results were subjected to analysis of variance using Assistat 7.7 beta (Silva and Azevedo, 2009). Treatment variances were initially assessed for homogeneity with Bartlett’s test. All variables showed homogeneous variances, and the effects of the treatments were tested with the F test. Means were compared with the Tukey test at 5%.

### RESULTS AND DISCUSSION

#### Plant growth

G12 and G4 had the highest LDW of all the genotypes (Tab. 3). G12, G8 and G4 were equal for TDW, all of which were higher than G9. There were no statistical
differences among the genotypes in terms of SDW yield (Tab. 3). G12, G8 and G4 were equal for main stem length, all of which were greater than G9 (Tab. 3). G8 had the highest number of secondary branches and tertiary branches of all the genotypes.

LDW yield becomes more relevant as glycosides are found in greater amounts in leaves (Jarma et al., 2010). LDW yields ranged from 349.40 kg ha⁻¹ (G9) to 836.40 kg ha⁻¹ (G12) with an overall mean of 621.15 kg ha⁻¹. These yields were lower than those found by Hastoy et al. (2019), Parris et al. (2016), Serfaty et al. (2013) and Espita et al. (2009).

In evaluating the same genotypes used in this study, Francisco (2015) found LDW yields of 4031.8 (G9) and 3733.1 kg ha⁻¹ (G12) in two harvests in the first year of cultivation. The lower yields found in this study could be explained by the genotype-environment interaction, where the photoperiod conditions observed by Francisco (2015) were favorable to genotype growth. Reduced yield is associated with the reduction of the vegetative cycle, which is caused by differences in seedling transplantation time (carried out in early December). The vegetative cycle in our study was 43 d shorter than the cycles of other studies, which affected the yields of the genotypes.

The seedlings of the stevia genotypes used in this study were produced by rooting branches. Stevia seedlings are typically produced from seeds (Lima Fiho, 2004). This enables the production and early transplanting of seedlings, which are more conducive to good crop growth (Ceunen and Geuns, 2013; Yoneda et al., 2017).

Late planting in this study reduced the crop growth period as a result of decreased photoperiod. As stevia is a short-day crop, it prematurely enters the reproductive phase, reducing the productive capacity of the plant (Ceunen and Geuns, 2013).

The variable LDW can also be influenced by planting density (Munz et al., 2018). The higher yields found by Daza et al. (2015), >3,500 kg ha⁻¹ and Espita et al. (2009), 1378 kg ha⁻¹, were a result of the use of more adapted genotypes and plant populations. The highest LDW yield found in G12 was associated with the highest LAI (Tab. 4), and there was a positive correlation between the variables (Tab. 5). A higher LAI increases the interception of photosynthetically active radiation and stimulates crop growth and photosynthetic production (Kumar et al., 2014).

Leaf dry weight (LDW), number secondary branches (secondary branches), number tertiary branches (tertiary branches), total leaf area (TLA), leaf area index (LAI), leaf area ratio (LAR), specific leaf area (SLA), rebaudioside A content (Reb A).

The TDW was influenced by LDW yield, with a positive but not significant correlation between the

<table>
<thead>
<tr>
<th>Genotypes</th>
<th>TLA (cm²)</th>
<th>LAI (m² m⁻²)</th>
<th>LAR (cm² g⁻¹)</th>
<th>SLA (cm² g⁻¹)</th>
<th>LWR (g g⁻¹)</th>
</tr>
</thead>
<tbody>
<tr>
<td>G4</td>
<td>719.93 b</td>
<td>5.75 b</td>
<td>42.80 a b</td>
<td>76.67 b</td>
<td>0.56 a</td>
</tr>
<tr>
<td>G8</td>
<td>428.01 b c</td>
<td>3.42 b c</td>
<td>23.49 b</td>
<td>63.41 b</td>
<td>0.37 a</td>
</tr>
<tr>
<td>G9</td>
<td>284.80 c</td>
<td>2.27 c</td>
<td>28.46 b</td>
<td>65.31 b</td>
<td>0.43 a</td>
</tr>
<tr>
<td>G12</td>
<td>1236.72 a</td>
<td>9.89 a</td>
<td>53.74 a</td>
<td>117.97 a</td>
<td>0.45 a</td>
</tr>
<tr>
<td>SE</td>
<td>101.14</td>
<td>0.68</td>
<td>3.66</td>
<td>6.53</td>
<td>0.02</td>
</tr>
</tbody>
</table>

Means followed by different letters indicate significant statistical differences according to the Tukey test at (P≤0.05) (n=4). SE, standard error.

<table>
<thead>
<tr>
<th>Indexes</th>
<th>TLA</th>
<th>LAR</th>
<th>SLA</th>
<th>LAI</th>
<th>Secondary branches</th>
<th>Tertiary branches</th>
<th>LDW</th>
<th>Reb A (mg g⁻¹)</th>
</tr>
</thead>
<tbody>
<tr>
<td>LDW</td>
<td>0.85**</td>
<td>0.68**</td>
<td>0.62**</td>
<td>0.85**</td>
<td>-0.31NS</td>
<td>0.11NS</td>
<td>0.91**</td>
<td></td>
</tr>
<tr>
<td>TDW</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>0.70**</td>
<td></td>
</tr>
</tbody>
</table>

NS = not significant; ** significant correlation at 1%.
variables (Tab 5). G9 had a TDW yield of 845.20 kg ha⁻¹, which was equal to G8 and G4, but lower than G12 (Tab. 3). These values are explained by the reduced TLA and LAI of the genotypes (Tab. 4).

There was a low correlation between the LDW yield and number of branches, with a negative, non-significant correlation between the LDW and number of secondary branches (Tab. 5). G8 had the highest number of lateral branches of all the genotypes (Tab. 3). According to Pal et al. (2015), the number of branches is associated with a longer growth period of the root system, where cytokines produced at the root apices and radicles stimulate lateral branches.

G8 had a longer production cycle than the other genotypes, and the harvest time occurred at 85 d after transplanting, which was also observed by Francisco (2015). G8, grown in the region of Curitiba, showed harvest conditions 121 d after transplanting (Francisco, 2015).

G12 (Tab. 4) showed the highest TLA, LAI and SLA of all the genotypes and an LAR equal to that of G4, and no differences for LWR were found among genotypes in the means separation test; a similar behavior was found by Francisco (2015). The highest TLA value in G12 was due to the highest LDW value, and there was a high correlation between LDW yield and TLA (Tab. 5).

According to Ceunen and Geuns (2013), LDW yield is directly influenced by the duration of the photoperiod. Longer photoperiods promote greater leaf expansion. An increase in leaf expansion favors total photosynthetic potential, promotes increased production of photoassimilates, and facilitates the partitioning of dry weight to other plant organs.

The genotypes assessed in this study were subjected to the same photoperiod. There was a decreasing photoperiod during the experiment period (Tab. 1), which, according to Francisco (2015) and Ceunen and Geuns (2013), is not favorable for the growth of stevia.

SLA is the ratio of leaf area to LDW. According to Poorter and Garnier (1999), it is an important physiological index representing leaf biomass allocation per unit of area. SLA is important for evaluating stevia genotypes because it represents the efficiency in synthesizing LDW, and the leaves present a higher concentrations of stevioside and rebadioside A (Jarman et al., 2010). G12 had the highest SLA value of all the genotypes (Tab. 4), which demonstrates that G12 had an increase in the expansion of the surface of the leaf blade, providing the development of thinner and more slender leaves. In our study, the SLA values were lower than those found by Francisco (2015), where values ranged from 10.82 to 17.37 m² kg⁻¹. These values are explained by the longer growth period in Francisco’s study (2015).

According to Magalhães (1979), LAR is a measure of the size of the photoassimilatory apparatus and serves as a variable to assess the effects of genotype, climate and management. G12 and G4 need a larger leaf area to synthesize 1 g of DW (Tab. 4). Francisco (2015) reported that genotypes with a low LAR may be associated with genotypes with a higher potential for stevioside and rebadioside A yield. Thus, G9 and G8 showed a higher photosynthetic efficiency compared to other genotypes (Tab. 4). The high LAR values in G12 and G4 are explained by the high TLA, which influences SLA, and increased SDW yield, showing a high correlation with TDW (Tab. 4).

According to Magalhães (1979), LAR is a measure of the size of the photoassimilatory apparatus and serves as a variable to assess the effects of genotype, climate and management. G12 and G4 need a larger leaf area to synthesize 1 g of DW (Tab. 4). Francisco (2015) reported that genotypes with a low LAR may be associated with genotypes with a higher potential for stevioside and rebadioside A yield. Thus, G9 and G8 showed a higher photosynthetic efficiency compared to other genotypes (Tab. 4). The high LAR values in G12 and G4 are explained by the high TLA, which influences SLA, and increased SDW yield, showing a high correlation with TDW (Tab. 4).
Stevioside and rebaudioside A yield

G12 showed the highest stevioside content and yield (Tab. 6). In terms of rebaudioside A, G8 was equal to G9, both of which were higher than the other genotypes (Tab. 6). G8 had the highest Reb A: St ratio and rebaudioside A yield (Reb A kg ha⁻¹).

LDW is influenced by the photoperiod (Ceunen and Geuns, 2013). Under increasing photoperiod conditions, stevia exhibits an increased LDW accumulation (Ceunen and Geuns, 2013). The genotypes in this study were exposed to a decreasing photoperiod, which indicates that the differences were associated with genetic differences.

The rebaudioside A content in G8 and G9 was equal and higher than those found in the other genotypes (Tab. 6). The values ranged from 20.39 mg g⁻¹ (2.03%) in G4 to 64.77 mg g⁻¹ (6.47%) in G8, which were lower than those of Francisco (2015).

The differences between our results and those of Francisco (2015) regarding rebaudioside A content are explained by the reduction of the crop cycle. Francisco (2015) reported that the genotypes showed a 27.9% reduction in stevioside accumulation and a 36.6% reduction in rebaudioside A accumulation under decreasing photoperiod conditions.

The correlation analysis between the morphophysiological characteristics and yield components of stevioside and rebaudioside A showed that the morphophysiological variables showed a high correlation with the stevioside content and yield, while the rebaudioside A content, Reb A: St ratio and rebaudioside A yield were negatively correlated with the LDW, TLA, LAI, SLA and LAR (Tab. 7).

The low rebaudioside A yield had a negative correlation with the LDW and TLA, LAI, SLA and LAR (Tab. 7). This negative correlation may explain the lower production of rebaudioside A in genotypes

<table>
<thead>
<tr>
<th>Genotype</th>
<th>Stevioside-St (mg g⁻¹)</th>
<th>Rebaudioside A-Reb A (mg g⁻¹)</th>
<th>Reb A: St ratio (g g⁻¹)</th>
<th>St yield (kg ha⁻¹)</th>
<th>Reb A yield (kg ha⁻¹)</th>
</tr>
</thead>
<tbody>
<tr>
<td>G4</td>
<td>176.97 b</td>
<td>20.39 b</td>
<td>0.11 c</td>
<td>133.66 b</td>
<td>15.33 c</td>
</tr>
<tr>
<td>G8</td>
<td>145.93 c</td>
<td>64.77 a</td>
<td>0.44 a</td>
<td>79.25 c</td>
<td>35.14 a</td>
</tr>
<tr>
<td>G9</td>
<td>128.47 c</td>
<td>49.05 a</td>
<td>0.38 b</td>
<td>44.72 d</td>
<td>17.12 c</td>
</tr>
<tr>
<td>G12</td>
<td>200.07 a</td>
<td>26.81 b</td>
<td>0.13 c</td>
<td>167.28 a</td>
<td>22.41 b</td>
</tr>
<tr>
<td>SE</td>
<td>7.64</td>
<td>4.87</td>
<td>0.04</td>
<td>12.67</td>
<td>2.06</td>
</tr>
</tbody>
</table>

Means followed by different letters indicate significant statistical differences according to the Tukey test at \(P \leq 0.05\) \((n = 4)\). SE: Standard error. The results of the stevioside content (Tab. 6) ranged from 128.47 mg g⁻¹ (12.8%) in G9 to 200.07 mg g⁻¹ (20%) in G12. These values were consistent with Francisco (2015) (stevioside content of 12% in G9 and 14.2% in G12). The stevioside content and yield were highly correlated with LDW (Tab. 7). According to Francisco (2015), genotypes that present a high LDW yield also present a high stevioside yield.

<table>
<thead>
<tr>
<th>Indexes</th>
<th>LDW</th>
<th>TLA</th>
<th>LAI</th>
<th>SLA</th>
<th>LAR</th>
</tr>
</thead>
<tbody>
<tr>
<td>St (mg g⁻¹)</td>
<td>0.98**</td>
<td>0.97*</td>
<td>0.97*</td>
<td>0.88**</td>
<td>0.93**</td>
</tr>
<tr>
<td>Reb A (mg g⁻¹)</td>
<td>-0.73NS</td>
<td>-0.70NS</td>
<td>-0.70NS</td>
<td>-0.63NS</td>
<td>-0.88NS</td>
</tr>
<tr>
<td>St (kg ha⁻¹)</td>
<td>0.98**</td>
<td>0.90**</td>
<td>0.90**</td>
<td>0.71**</td>
<td>0.78**</td>
</tr>
<tr>
<td>Reb A (kg ha⁻¹)</td>
<td>0.09NS</td>
<td>-0.08NS</td>
<td>-0.08NS</td>
<td>-0.63**</td>
<td>-0.39**</td>
</tr>
<tr>
<td>Reb A:St ratio</td>
<td>-0.31NS</td>
<td>-0.80NS</td>
<td>-0.80NS</td>
<td>-0.71NS</td>
<td>-0.93NS</td>
</tr>
</tbody>
</table>

** Significant correlation at 1%, * significant correlation at 5%, NS not significant.

Leaf dry weight (LDW), total leaf area (TLA), leaf area index (LAI), specific leaf area (SLA), leaf area ratio (LAR), stevioside content (St mg g⁻¹), rebaudioside A content (Reb A mg g⁻¹), stevioside yield (St kg ha⁻¹), rebaudioside A yield (Reb A kg ha⁻¹), rebaudioside A: stevioside ratio (Reb A: St ratio).
with higher growth rates. According to Bondarev et al. (2010), the smaller leaves in stevia present an increased density of glandular trichomes and, consequently, a higher rebaudioside A content.

The reb A: St ratio is important for selecting stevia genotypes. According to Mota et al. (2011), the ideal Reb A: St ratio for industrial use is 1. The genotypes did not show the desired Reb A: St ratio. G8 had the best Reb A: St ratio of all the genotypes (Tab. 6). The ratios found in this study were lower than those observed by Francisco (2015), Tavarini et al. (2015) and Mandal et al. (2013).

G12 showed the highest stevioside yield (Tab. 6). Genotypes with a higher LDW yield and higher secondary metabolite concentration (Tavarini et al., 2015; Vasilakoglou et al., 2016) tend to have better stevioside yield. There is a strong correlation between LDW yield and TLA, LAI, SLA and LAR (Tab. 7) for stevioside yield.

CONCLUSIONS

The G8 genotype had the highest rebaudioside A content, the highest rebaudioside A yield and the best rebaudioside A: stevioside ratio under the productive conditions with a decreasing photoperiod in the Alto Vale do Itajaí region (SC).

The G12 genotype showed the best growth rate, morphophysiological indexes, stevioside content and yield.

No genotype evaluated in this study had a rebaudioside A: stevioside ratio that was suitable for industry requirements.

Conflict of interest: this manuscript was prepared and reviewed with the participation of all authors, who declare that there exists no conflict of interest that puts at risk the validity of the presented results.

BIBLIOGRAPHIC REFERENCES


