

Morphological characterization and hierarchical classification of 40 bush pea genotypes (*Pisum sativum* L.)

Caracterización morfológica y clasificación jerárquica de 40 genotipos de arveja arbustiva (*Pisum sativum* L.)



VERÓNICA ROSERO-LOMBANA¹
OSCAR CHECA-CORAL^{1, 2}

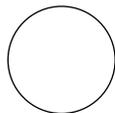
Sample of the variability of shape, color and size of bush pea seed from the collection of the Universidad de Nariño (Colombia).

Photo: O. Checa-Coral.

ABSTRACT

The Universidad de Nariño is home to a collection of bush pea plantlets that are a source of biodiversity for the genetic improvement of pea species in Colombia. The characterization of these accessions is required to identify genotypes with attributes that could be used in the search for new varieties. For the morphological characterization, 40 pea accessions were planted in Pasto, Colombia. 23 quantitative variables and 12 qualitative variables were documented, descriptors proposed for this species by the European Union in 2003. The data were subjected to Principal Component Analysis and Multiple Correspondence Analysis. Finally, a hierarchical classification method was applied using Ward's method. The first four components, which explained 78.80% of the total variability of the population, were selected for the quantitative variables. Four groups were identified. Genotypes with the *afila* gene, which are of interest for pea breeding programs, were found in groups 1 and 2. The highest seed weight was in group 1, and the genotypes with the best reaction to powdery mildew were in group four. For the qualitative variables, the first six factors, which described 60.51% of the variability, were selected, and the hierarchical classification analysis resulted in five groups. The qualitative characteristics that contributed more to the differentiation of the groups included leaflet type, hilum color, degree of curvature of the pod, color and shape of the grain.

Additional key words: grain legumes; biological collections; selection; classification; principal component analysis; multiple correspondence analysis; variability.



¹ Universidad de Nariño, Faculty of Agricultural Sciences, Pasto Campus, Grupo de Investigación Cultivos Andinos, San Juan de Pasto (Colombia). ORCID Rosero-Lombana, V.: 0000-0002-3262-7073; ORCID Checa-Coral, O.: 0000-0002-6929-7717

² Author for correspondence. cicagrarias@hotmail.com

RESUMEN

La Universidad de Nariño cuenta con una colección de arvejas arbustivas que constituyen una fuente de biodiversidad para el mejoramiento genético de esta especie en Colombia. La caracterización de las accesiones es requerida para identificar genotipos con atributos que sean de interés para el proceso de obtención de nuevas variedades. Con el objeto de realizar la caracterización morfológica, 40 accesiones de arveja arbustiva fueron sembradas en Pasto-Colombia, para registrar 23 variables cuantitativas y 12 variables cualitativas que corresponden a los descriptores propuestos para esta especie por la Unión Europea en el 2003. Los datos obtenidos se sometieron a Análisis de Componentes Principales y Análisis de Correspondencias Múltiples. Finalmente, se aplicó el método de clasificación jerárquica utilizando el criterio de Ward. Para las variables cuantitativas se seleccionaron los cuatro primeros componentes que explicaron el 78,80% de la variabilidad total de la población. Se identificaron cuatro grupos. Los genotipos con el gen afila que son de interés para el mejoramiento de arveja se encontraron en los grupos 1 y 2. El mayor peso de semilla se encontró en el grupo 1 y los genotipos con mejor reacción al mildiú polvoroso en el grupo cuatro. Para las variables cualitativas se seleccionaron los primeros seis factores que describen el 60,51% de la variabilidad y el análisis de clasificación jerárquica resultó en la creación de cinco grupos. Las características cualitativas que más contribuyeron a la diferenciación de los grupos fueron tipo de folíolos, color del hilio, curvatura de la vaina, color y forma del grano.

Palabras clave adicionales: leguminosas de grano; colecciones biológicas; selección; clasificación; análisis de componentes principales; análisis de correspondencias múltiples; variabilidad.

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INTRODUCTION

The pea is one of the older cultivated plants used by mankind. Wild pea fields can still be found today in Afghanistan, Iran and Ethiopia (Espósito *et al.*, 2007). Broad genetic diversity has been reported in peas by Bhuvanewari *et al.* (2017) and Singh *et al.* (2017). This diversity has been conserved in gene banks and studied for the last 20 years (Singh *et al.*, 2013; Warkentin *et al.*, 2015). 72,000 pea accessions are preserved in 25 large collections, and 27,000 accessions make up part of 146 collections, including various wild relatives (Smykal *et al.*, 2011; Smykal *et al.*, 2013; Smykal *et al.*, 2014; Trněný *et al.*, 2018). *Pisum sativum* L. is used for human consumption as a dry seed or as fresh vegetables grown from immature grains (Ma *et al.*, 2017). Dried peas are grown during the summer in Australia, Canada, China, India, Eastern, Europa and the northwest United States (Wu, *et al.*, 2017). Genetic diversity is the main input for the genetic improvement of a cultivated species; therefore, it is a prerequisite for any improvement program to increase yields and stabilize production in the face of disease epidemics and fluctuation of environmental conditions. (Abubakkar *et al.*, 2011; Gatti *et al.*, 2011; Tiwary and Lavanya, 2012). Local and wild varieties are maintained in germplasm banks as genetic

resources; however, their use has been limited when breeding for yield (Ali *et al.*, 2007). Since inheritance acts as a predictive tool to express the reliability of phenotypic traits, high heritability could help in the effective selection of particular characteristics and when designing future selection programs (Mallu *et al.*, 2014).

Morphological characterization is the first step in describing and classifying germplasm (Bouhadida *et al.*, 2013). Understanding morphological characteristics facilitates the identification and selection of desirable attributes, design of new populations, gene transfer, and resistance to biotic and abiotic factors (Singh *et al.*, 2014). In the characterization process, traits with high heritability are favorable because this implies little environmental influence. This ensures that the genotype environment interaction has a reduced effect on variables that characterize different accessions.

The objective of the present investigation was to carry out a morphological characterization of 40 bush pea accessions from European and South American countries, donated to the Universidad de Nariño by

the Corporación Colombiana para la Investigación Agropecuaria (Agrosavia) and the Universidad Nacional de Colombia or grown from plants through local crosses to identify and classify genotypes and attributes of interest for the genetic improvement of species in Colombia.

MATERIALS AND METHODS

Location. This research was carried out in 2018 and 2019 at the Centro Internacional para Produccion Limpia “LOPE Spanish traducción”, which belongs to the Servicio Nacional de Aprendizaje of Colombia (SENA), located in the municipality of Pasto, Department of Nariño, with an altitude of 2,612 m a.s.l., at 01°12'28" N and 77°15'06" W, with an average temperature of 14°C, annual rainfall of 841 mm and relative humidity of 73%

Genetic material. 40 bush pea (*Pisum sativum* L.) genotypes belonging to the Andean Crops Research Group (GRICAND) of the Faculty of Agricultural Sciences at the Universidad de Nariño were used in this study. In the last decade, GRICAND developed a collection of bush pea genotypes of different origins, donated by the Universidad Nacional de Colombia, the Agrosavia germplasm bank, and the improvement program of the Universidad de Nariño. The sowing was carried out in 2018; 40 lines were sown in a block with plots of three two-meter rows, with 0.5 m between rows and 0.1 m between sites, depositing one seed per site. The land preparation and agronomic management of the crop was done according to the recommendations of Buitrago *et al.* (2006).

Morphological characterization. The 40 bush pea genotypes were characterized with 12 qualitative and 23 quantitative variables, using the descriptors proposed by the European Union (2003). In addition, the quantitative variables included the percentage reaction to powdery mildew, and the readings were taken with the assessment scales proposed by Checa and Rodríguez (2015). In each of the accessions, five plants in the middle furrow were selected, where the data for the variables were recorded. For the quantitative variables, the results were recorded from the average of the measurements, and, for the qualitative variables, the observations were recorded.

Statistical analysis. The results for the quantitative variables were subjected to principal component analysis (PCA). The quantitative variables were

selected, discarding or eliminating those that resulted in a coefficient of variation of less than 20%.

The variables selected for the PCA were: W100 = Weight of 100 seeds; Pppl = Pods per plant; Bbr = Basal branches; Sbr = Secondary branches; Pmb = Peduncle in main branch; Psbr = Peduncle secondary branch; Nle = Number of leaflets; Sl = Stipule length; Sw = Stipule width; Lw = Leaflet width; Ll = Leaflet length; and Pmldr = Reaction to powdery mildew.

A multiple correspondence analysis (MCA) was used for the qualitative variables. Ward's distances were used in the classification and establishment of grouping levels and for the construction of dendrograms. The data were processed using Spad v3.5.

RESULTS AND DISCUSSION

Principal component analysis

Four principal components explained 78.80% of the variability between the analyzed accessions (Tab. 1). The first component explained 44.83% of the total observed variability. The variables that contributed most to this component were number of leaflets (Nle), leaflet width (Lw), leaflet length (Ll) and Peduncle secondary branch (Psbr). The second component explained 16.53% of the total observed variability, where the variables number of basal branches (Bbr) and Leaflet length (Ll) had a notable contribution. The third component explained 9.39% of the total variability. The variables that contributed more to the third component were stipule width (Sw) and resistance to powdery mildew (Pmldr). Finally, the fourth component explained 8.05% of the total variability, where the variable resistance to powdery mildew (Pmldr) contributed most (Tab. 1 and 2).

The results agree with the report by Gixhari *et al.* (2014), who stated that the weight of 100 seeds, the width of the leaflet, and the length of the leaflet are among the characteristics that explain much of the variability in peas. They also agree with Hanci and Cebeci (2018), who included the width and length of the leaflet and the length and width of the stipule among the variables with significant contribution to the main components.

Table 1. Eigenvalues and percentages of variability explained by the principal components.

Principal component	Eigenvalues	Percentage	Total percentage
1	5.3790	44.83	44.83
2	1.9836	16.53	61.36
3	1.1269	9.39	70.75
4	0.9665	8.05	78.80
5	0.7617	6.35	85.15
6	0.6178	5.15	90.30
7	0.4542	3.79	94.08
8	0.3182	2.65	96.73
9	0.1810	1.51	98.24
10	0.1604	1.34	99.58
11	0.0472	0.39	99.97
12	0.0035	0.03	100.00

Table 2. Contribution of the quantitative variables to the conformation of the principal components.

Variables/Components	1	2	3	4
C2 – W100 (G)	-0.67	-0.27	-0.32	0.00
C3 – PPPL	0.64	0.02	0.06	-0.20
C4 – BBR	0.18	-0.79	-0.28	0.12
C5 – SBR	0.66	-0.23	0.02	0.32
C6 – PMB (CM)	0.52	-0.46	0.36	-0.47
C7 – PSBR (CM)	0.80	-0.21	0.37	0.01
C8 – NLE	0.94	0.24	-0.09	0.04
C9 – (SL) (CM)	0.17	-0.79	-0.06	0.42
C10 – (SW) (CM)	-0.55	-0.38	0.61	-0.15
C11 – (LW) (CM)	0.95	0.14	-0.04	0.06
C12 – (LL) (CM)	0.95	0.20	-0.06	0.09
C13 – PMILDR%	-0.32	0.31	0.54	0.62

Four groups were established with the Classification Analysis dendrogram generated from the PCA (Fig. 1), as shown in table 3. The first group included 15 genotypes, (ARB012, ARB014, ARB013, ARB021, ARB016, ARB005, ARB011, ARB007, ARB008, ARB010, ARB009, ARB004, ARB002, ARB003 and ARB001) 37.5% of the studied population. This group in particular stood out for the weight values per 100 kernels (74.87 g) when compared to the overall average (65.20 g). Other characteristics in this group were recorded below the general average.

The presence of zero values in the number category indicated that the genotypes of this group were *afila*, characterized by a lack of leaflets as the result of the recessive *af* gene, which replaces leaves with tendrils (Amin *et al.*, 2010).

Weight of 100 grains is an important variable because it represents the part of the legume that is useful for consumption. In general, the varieties with a high grain weight per pod have a high market value. The genotypes of this group could contribute to the improvement of this characteristic, but genetic improvement has compensation mechanisms for the components of yield. When the number of seeds per pod is increased, the number of pods per plant and the seed weight could be reduced (Tiemerman *et al.*, 2005). Gupta *et al.* (1984) reported that the expression of seed weight is governed by additive and non-additive genetic effects. However, the selection genotypes with more pods per plant and a greater number of seeds per pod could improve yield (Brijendra *et al.*, 2013; Iqbal *et al.*, 2017; Kumar *et al.*, 2017).

Group two was made up of nine individuals: ARB017, ARB015, ARB024, ARB018, ARB020, ARB019, ARB023, ARB022 and ARB006, which corresponded to 22.5% of the population, and had the widest stipule (12.72 cm), with respect to the overall average (9.86 cm). The expression was regulated by the presence of the *lat* gene (Amin *et al.*, 2010). The zero values for the number, width and length of foliole indicated that this group had *afila* genotypes. Unlike the genotypes in group one, the stipules were large, giving them the ability to compensate for the foliar area that is decreased when lateral leaves are replaced by tendrils.

Group three was made up of the individuals ARB031, ARB025, ARB029, ARB038, ARB040, ARB027, ARB026, ARB028, ARB030 and ARB039, which corresponded to 25% of the total population. The accessions were characterized by an above-average foliole length, number of leaflets and foliole width and a below-average weight of 100 grains and stipule width.

Group four had six individuals (ARB033, ARB034, ARB032, ARB035, ARB037 and ARB036), which represented 15% of the total population. The variables that were above the overall average and characterized this group were size of peduncle in the main branch, foliole width, size of peduncle in the secondary branch, number of leaflets, leaflet length, number of secondary branches, number of basal branches, and stipule length.

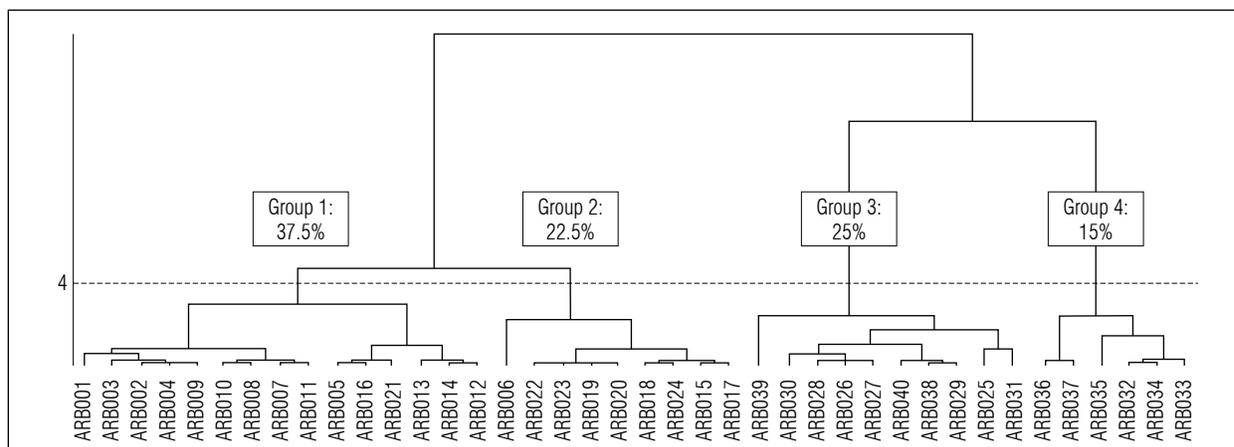


Figure 1. Dendrogram analysis of quantitative variables classification.

This group included genotypes with the densest foliage, requiring an adjustment in population density to obtain better results. The lines of this group were less susceptible to powdery mildew. *Erysiphe pisi* (Ep) is the main causal agent of powdery mildew

(PM) in peas and is responsible for 25-80% losses in yield worldwide (León *et al.*, 2020; Nisar *et al.*, 2006). The powdery mildew decreases pod quality and is a yield-limiting factor (Nag and Khare, 2017; Sharma *et al.*, 2017).

Table 3. Characterization with continuous variables in groups or categories.

Probability	Average		Standard deviation		Variables
	Group	General	Group	General	
Group 1					
0.001	74.87	65.20	9.32	15.14	W100 (g)
0.007	8.80	9.83	0.96	2.03	Pmb (cm)
0.000	1.51	2.35	0.82	1.19	Sbr
0.000	20.39	27.21	6.92	9.39	Pppl
0.000	0.00	2.03	0.00	2.58	Nle
0.000	0.00	1.28	0.00	1.61	LI (cm)
0.000	0.00	2.00	0.00	2.48	LI (cm)
0.000	3.64	5.70	1.28	2.46	Psbr (cm)
Group 2					
0.000	12.72	9.86	0.82	2.08	Sw (cm)
0.004	0.00	2.03	0.00	2.58	Nle
0.004	0.00	1.28	0.00	1.61	Lw (cm)
0.003	0.00	2.00	0.00	2.48	LI (cm)
Group 3					
0.000	4.98	2.00	0.54	2.48	LI (cm)
0.000	4.98	2.03	0.71	2.58	Nle
0.000	3.03	1.28	0.55	1.61	Lw (cm)
0.002	52.90	65.20	17.08	15.14	W100 (g)
0.000	7.73	9.86	1.02	2.08	Sw (cm)

Continued

Continuation Table 3. Characterization with continuous variables in groups or categories.

Probability	Average		Standard deviation		Variables
	Group	General	Group	General	
Group 4					
0.000	13.50	9.83	0.86	2.03	Pmb (cm)
0.000	3.52	1.28	0.42	1.61	Lw (cm)
0.000	9.03	5.70	1.12	2.46	Psbr (cm)
0.001	5.22	2.03	1.50	2.58	Nle
0.001	5.07	2.00	0.59	2.48	Ll (cm)
0.001	3.75	2.35	1.05	1.19	Sbr
0.004	2.28	1.74	0.38	0.53	Bbr
0.008	6.63	5.47	1.26	1.26	Sl (cm)
0.000	0.36	0.71	0.37	0.27	Pmldr (%)

Multiple correspondence analysis

Nine qualitative traits that showed variability (Dgrc, Typc, Scol, Ssur, Hilc, Hab, Tleaf, Mf, Lmar) showed that the first factor explained 18.99%, the second one explained 11.25%, the third explained 8.56%, the fourth explained 7.98, and the fifth explained 7.72%. These first five factors explained 54.49% of the total cumulative variability (Tab. 4).

The analysis of the effects of the qualitative variables on the factorial axes (Tab. 5) indicated that the variables with the highest contribution to the first factor (factor one) were grain shape (Sseed = 18.0%), margin of foliole (Lmar = 17.5%), seed surface (Ssur = 16.2%), leaf type (Tleaf = 14.7%), and degree of curvature of the pod (Dgrc = 13.1%). The second factor was mainly defined by grain surface (Ssur = 20.8%), leaf margin (Lmar = 18.2%), hilum color (Hilc = 17.8%), and seed shape (Sseed = 17.2%). For the third factor, the notable, contributing variables were grain color (Scol. = 25.5%), type of pod curvature (Typc = 23.6%), growth habitat (Hab = 13.8%) and leaflet margin (Lmar = 11.6%). In the fourth factor, the variables with the highest contribution were leaflet margins (Lmar = 29.1%) and type of pod curvature (Typc = 27.9%). For the fifth factor, the variables with the highest participation were grain shape (31.5%), pod curvature type (Typc = 23%), and seed surface (Ssur = 20.7%).

According to the hierarchical classification analysis, the 40 lines of pea plants were organized into five groups (Fig. 2, Tab. 6). The first group had 42.5% of the individuals (ARB008, ARB007, ARB020, ARB006,

Table 4. Eigenvalues and percentages of variability explained by the factors resulting from the multiple correspondence analysis (MCA).

Factor	Eigenvalues	Percentage	Accumulated percentage
1	0.4852	18.99	18.99
2	0.2874	11.25	30.23
3	0.2188	8.56	38.79
4	0.2039	7.98	46.77
5	0.1972	7.72	54.49
6	0.1539	6.02	60.51
7	0.1352	5.29	65.81
8	0.1218	4.76	70.57
9	0.1188	4.65	75.22
10	0.0988	3.87	79.09
11	0.0977	3.82	82.91
12	0.0891	3.49	86.40
13	0.0732	2.86	89.26
14	0.0637	2.49	91.76
15	0.0575	2.25	94.00
16	0.0507	1.98	95.99
17	0.0376	1.47	97.46
18	0.0239	0.93	98.39
19	0.0180	0.70	99.10
20	0.0161	0.63	99.73
21	0.0055	0.22	99.94
22	0.0014	0.06	100.00
23	0.0000	0.00	100.00

ARB022, ARB010, ARB024, ARB021, ARB019, ARB023, ARB001, ARB002, ARB004, ARB017, ARB011, ARB018 and ARB ARB009). 100% of the individuals in this class had an *afila af* gene, which replaces leaves with tendrils.

The accessions that form this group are of special interest to the improvement of pea plants because they are *afila* genotypes. The *afila* gene is recessive and leads to phenotypic expression only if it occurs in a homozygous recessive combination (*afaf*); it transforms leaves into tendrils (Singh *et al.*, 2013). Because of the presence of the *afila* gene, plants resist early overturning, which affects varieties with conventional foliage; the greater the development of the tendril, the greater resistance to overturning (Wang *et al.*, 2003). In addition, greater aeration favors plants that are less affected by foliar diseases. (Checa *et al.*, 2020).

Table 5. Contribution of the qualitative variables to the factor axes in the analysis of multiple correspondences.

Variable	1	2	3	4	5
Dgrc	13.1	0.2	2.4	5.0	0.3
Typc	8.2	8.0	23.6	27.9	23.0
Scol	9.7	3.9	25.5	4.9	9.0
Ssur	16.2	20.8	7.9	5.1	20.7
Hilc	0.1	17.8	10.9	5.4	0.2
Hab	2.4	9.9	13.8	11.0	4.3
Tleaf	14.7	4.1	0.2	6.6	0.0
Mf	17.5	18.2	11.6	29.1	11.0
Lmar	18.0	17,2	4.1	5.1	31.5

In restrictive humidity, these *afila* varieties tend to increase productivity in comparison to non-*afila* varieties (Jannink *et al.*, 1996; Mihailovic, 2008).

All the individuals in this class presented a hyaline hilum color. In addition, 88.24% of the individuals in this group presented a rectangular shape with several shapes (C13 = 4). Another characteristic that contributed to this group was growth habitat; 47.06% were semi-twining individuals (C10 = 2).

Group two was made up of eight individuals, representing 20% of the population: ARB012, ARB016, ARB040, ARB005, ARB015, ARB013, ARB014, and ARB003. In this group, 87.50% had a light brown hilum, and 12.50% had a hilum. There were no qualitative attributes of great interest in this group for the genetic improvement of this species.

The third group was made up of seven individuals that represented 17.5% of the studied population. 100% of these genotypes had normal leaves or tendrils, and 85.71% had rough seed surfaces. The wrinkled-seeded phenotype has been associated with natural or induced mutations that impact concentrations of starch and sucrose (Santos *et al.*, 2019).

Group four had four individuals that represented 10% of the population; this group was characterized by a degree of curvature in the soft sheath (G2 = 2). This characteristic confers high resistance for transport, as compared to pods with a strong degree of curvature. In addition, all individuals in this group presented a green grain color (C4 = 4) and 75% smooth seed surface (C5=1).

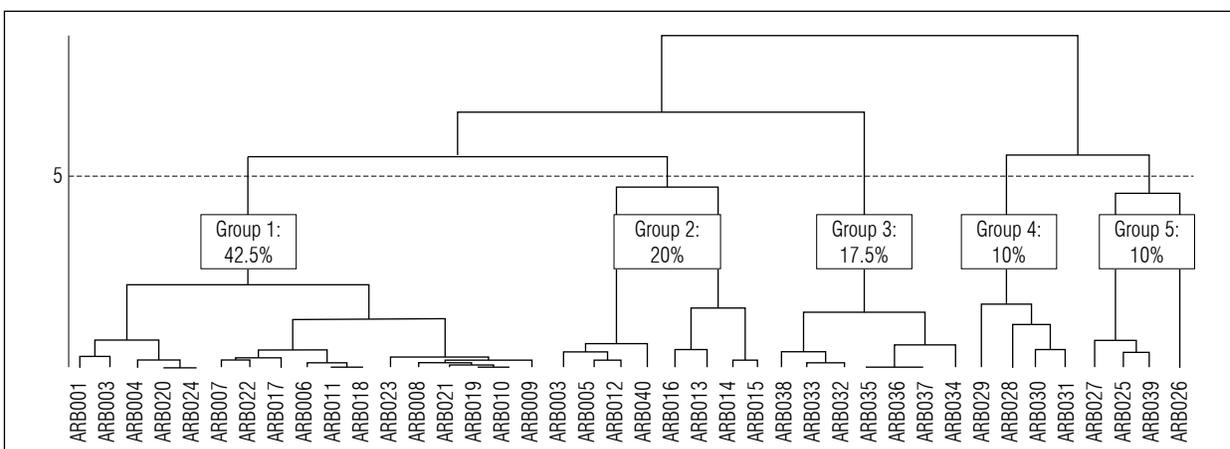


Figure 2. Dendrogram classification analysis for qualitative variables.

The fifth group was made up of four individuals that represented 10% of the population. All individuals in this group had round seeds; 75% had a cream-colored, and 75% were smooth, which implied a predominance of the *R Rb* genotype. The predominance

of the double dominant *Pla Qua*, which indirectly determines rounded seed shapes, is also expected, favoring non-compact arrangements of seeds in pods (UPOV, 1994).

Table 6. Characterization of group categories.

Probability	Mean		Std. deviation		Characteristic variables	Weight
	Group	Overall	Group	Overall		
Group 1						
0.000	70.83	100	60	C12=1	Lmar	24
0.000	70.83	100	60	C11=3	Tleaf	24
0.003	54.84	100	77.5	C6=1	Hilc	31
0.009	57.69	88.24	65	C13=4	Sseed	26
0.009	100	29.41	12.5	C10=3	Hab	5
0.009	28.57	47.06	70	C10=2	Hab	28
0.003	0.00	0.00	22.5	C6=2	Hilc	9
0.000	0.00	0.00	40	C11=1	Tleaf	16
0.000	0.00	0.00	0.00	C11=2	Tleaf	0
Group 2						
0.000	77.78	87.5	22.5	C6=2	Hilc	9
0.000	3.23	12.5	77.5	C6=1	Hilc	31
0.000	0	0	0	C11=2	Tleaf	
Group 3						
0.001	43.75	100	40	C11=1	Tleaf	16
0.003	46.15	85.71	32.5	C5=4	Ssur	13
0.001	0.00	0.00	60	C11=3	Tleaf	24
0.001	0.00	0.00	60	C12=1	Lmar	24
0.000	0.00	0.00	0.00	C11=2	Tleaf	0
Group 4						
0.001	50	100	20	C2=2	Dgrc	8
0.008	50	75	15	C5=1	Ssur	6
0.008	100	50	5	C3=5	Tipc	2
0.008	100	50	5	C12=2	Lmar	2
0.008	30.77	100	32.5	C4=4	Scol	13
0.001	0.00	0.00	80	C2=1	Dgrc	32
0.000	0.00	0.00	0	C11=2	Tleaf	0
Group 5						
0.000	57.14	100	17.5	C13=1	Sseed	7
0.000	100	75	7.5	C4=1	Scol	3
0.008	50	75	15	C5=1	Ssur	6
0.00	0.00	0.00	0	C11=2	Tleaf	0

CONCLUSIONS

The quantitative variables (weight of 100 grains, stipulate size, width and length of leaflets, number of leaflets, and peduncle size on the main branch) and the qualitative variables (leaf type, hilum color, grain type, and degree of curvature of the pod), contributed to the variability of the pea collection characterized in this study.

Among the variables of interest for pea improvement, the *afila* trait conferred by the *af* gene was found in groupings one and two of the quantitative variables and in group one of the qualitative variables. The highest seed weight was found in the genotypes in grouping one of the quantitative variables, and the genotypes resistant to powdery mildew were in grouping four.

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Conflict of interests: The manuscript was prepared and reviewed with the participation of the authors, who declare that there exists no conflict of interest that puts at risk the validity of the presented results.

BIBLIOGRAPHY REFERENCES

- Abubakkar, M., N. Nobel, A. Ali, M. Ashraf, S. Niaz, and K. Mahmood. 2011. Characterization of pea germplasm. *Int. J. Veg. Sci.* 17(3), 246-258. Doi: 10.1080/19315260.2010.544380
- Ali, Z., A. Qureshi, W. Ali, H. Gulzar, M. Nisar, and A. Ghafoor. 2007. Evaluation of genetic diversity present in pea (*Pisum sativum* L.) germplasm base on morphological traits, resistance to powdery Mildew and molecular characteristics. *Pak. J. Bot.* 39(7), 27-39.
- Amin, A., F. Mushtaq, P.K. Singh, K.P. Wani, S. Spaldon, and N. Nazir. 2010. Genetics and breeding of pea. *Int. J. Curr. Res.* 10, 028-034.
- Bhuvaneshwari, S., S.K. Sharma, P. Punitha, K.S. Shashidhar, K.L. Naveenkumar, and N. Prakash. 2017. Evaluation of morphological diversity of field pea [*Pisum sativum* subsp. *arvense* (L.)] germplasm under sub-tropical climate of Manipur. *Legume Res.* 40, 215-223. Doi: 10.18805/lr.v0i0E.10756
- Bouhadida, M., F. Srarfi, I. Saadi, and M. Kharrat. 2013. Molecular characterization of pea (*Pisum sativum* L.) using microsatellite markers. *J. Appl. Chem.* 5(1), 57-61. Doi: 10.9790/5736-0515761
- Brijendra, K., K. Adesh., K. Ashutosh, and L. Roopa. 2013. Selection strategy for seed yield and maturity in field pea (*Pisum sativum* L. *arvense*). *Afr. J. Agric. Res.* 8(44), 5411-5415.
- Buitrago, J. and C. Duarte. 2006. El cultivo de arveja en Colombia. Fondo Nacional de Leguminosas, Bogota.
- Checa, O. and M. Rodríguez. 2015. Resistencia a oídio (*Erysiphe polygoni*) y rendimiento en arveja afila (*Pisum sativum* L.). *Temas Agrarios* 20(2), 58-71. Doi: 10.21897/rta.v20i2.759
- Checa, O.E., M. Rodriguez, X. Wu, and M.W. Blair. 2020. Introgression of the *Afila* gene into climbing garden pea (*Pisum sativum* L.). *Agronomy* 10, 1537. Doi: <https://doi.org/10.3390/agronomy10101537>
- Espósito, M., E. Martin, V. Cravero, and E. Cointry. 2007. Characterization of pea accessions by SRAP's markers. *Sci. Hortic.* 113(4), 329-335, Doi: 10.1016/j.scienta.2007.04.006
- European Union. 2003. Protocol for distinctness, uniformity and stability tests *Pisum sativum* L. *Sensu lato*, PEA, European Union Community Plant Variety Office, Brussels.
- Gatti, I., M.A. Espósito, P. Almirón, V.P. Cravero, and E.L. Cointry. 2011. Diversity of pea (*Pisum sativum*) accessions based on morphological data for sustainable field pea breeding in Argentina. *Genet. Mol. Res.* 10(4), 3403-3410. Doi: 10.4238/2011.October.31.8
- Gixhari, B., H. Vrap, and V. Hobdari. 2014. Morphological characterization of pea (*Pisum sativum* L.) genotypes stored in Albanian genebank. In: First International Conference "Biotechnology in Agriculture". Agricultural University of Tirana, Tirana.
- Gupta, K., R. Waldia, B. Dahiya, K. Singh, and D. Sood. 1984. Inheritance of seed yield and quality traits in peas (*Pisum sativum* L.). *Theor. Appl. Genet.* 69(2), 133-137. Doi: 10.1007/BF00272884
- Hanci, F. and E. Cebeci. 2018. Determination of morphological variability of different *Pisum* genotypes using principal component analysis. *Legume Res.* 42(2), 162-165. Doi: 10.18805/LR-438
- Iqbal, A., S. Shah, M. Nisar, and A. Ghafoor. 2017. Morphological characterization and selection for high yielding and powdery mildew resistant pea (*Pisum sativum*) lines. *Sains Malays.* 46(10), 1727-1734. Doi: 10.17576/jsm-2017-4610-08
- Jannink, J., M. Liebman, and L. Merrick. 1996. Biomass production and nitrogen accumulation in pea, oat and vetch green manure mixtures. *Agron. J.* 88(2), 231-240. Doi: 10.2134/agronj1996.00021962008800020019x

- Kumar, M., M.S. Jeberson, N.B. Singh, and R. Sharma. 2017. Genetic analysis of seed yield and its contributing traits and pattern of their inheritance in field pea (*Pisum sativum* L.). *Int. J. Curr. Microbiol. Appl. Sci.* 6, 172-181. Doi: 10.20546/ijcmas.2017.606.021
- León, D., Ó. Checa, and P. Obando. 2020. Inheritance of resistance of two pea lines to powdery mildew. *Agron. J.* 112(4), 2466-2471. Doi: 10.1002/agj2.20253
- Ma, Y., C.J. Coyne, M.A. Grusak, M. Mazourek, P. Cheng, D. Main, and R.J. McGee. 2017. Genome-wide SNP identification, linkage map construction and QTL mapping for seed mineral concentrations and contents in pea (*Pisum sativum* L.). *BMC Plant Biol.* 17(43), 1-17. Doi: 10.1186/s12870-016-0956-4
- Mallu, T.S., S.G. Mwangi, A.B. Nyende, N.V.P.R.G. Rao, D.A. Odeny, A. Rathore, and A. Kumar. 2014. Assessment of genetic variation and heritability of agronomic traits in chickpea (*Cicer arietinum* L.). *Int. J. Agron. Agric. Res.* 5(4), 76-88.
- Mihailovic, V. 2008. Componentes de rendimiento de grano afila (af) líneas de guisantes forrajeros (*Pisum sativum* L.). Instituto de Cultivos y Hortalizas, NoviSad, Serbia.
- Nag, U.K. and C.P. Khare. 2017. Epidemiological studies on powdery mildew of vegetable pea. *Plant Arch.* 17(1), 171-176.
- Nisar, M., A. Ghafoor, M.R. Khan, and A.S. Qureshi. 2006. Screening of *Pisum sativum* L. germplasm against *Erysiphe pisi* Syd. *Acta Biol. Crac. Ser. Bot.* 48(2), 33-37.
- Santos, C.S., B. Carbas, A. Castanho, M.W. Vasconcelos, M.C. Vaz Patto, C. Domoney, and C. Brites. 2019. Variation in pea (*pisum sativum* L.) seed quality traits defined by physicochemical functional properties. *Foods* 8(11), 570. Doi: 10.3390/foods8110570
- Sharma, R.L., T. Mishra, R. Bhagat, and V.K. Swarnkar. 2017. Comparative efficacy of different new fungicides against powdery mildew disease of field pea (*Pisum sativum* L.). *Int. J. Curr. Microb. Appl. Sci.* 6(4), 1349-1360.
- Singh, B., T. Chaubey, D. Upadhyay, A. Jha, and S. Pandey. 2014. Morphological characterization of vegetable pea (*Pisum sativum* L. spp. hortense) genotypes and their application for distinctiveness, uniformity and stability testing. *Indian Legum. Res.* 37(5), 547-551. Doi: 10.5958/0976-0571.2014.00674.2
- Singh, M., H. Upadhyaya, and I. Bisht. 2013. Genetic and genomic resources of grain legume improvement. Elsevier, London.
- Singh, S.R., N. Ahmed, D.B. Singh, K.K. Srivastva, R.K. Singh, and A. Mir. 2017. Genetic variability determination in garden pea (*Pisum sativum* L. sub sp. hortense Asch. and Graebn). by using the multivariate analysis. *Legum. Res.* 40, 416-422.
- Smýkal, P., C. Coyne, M. Ambrose, N. Maxted, H. Schaefer, and M. Blair. 2014. Legume crops phylogeny and genetic diversity for science and breeding. *Crit. Rev. Plant Sci.* 34, 43-104. Doi: 10.1080/07352689.2014.897904
- Smýkal, P., C. Coyne, R. Redden, and N. Maxted. 2013. Peas. In: Singh, M., H.D. Upadhyaya, and I.S. Bisht (eds.). Genetic and genomic resources of grain legume improvement. Elsevier, Amsterdam. Doi: 10.1016/B978-0-12-397935-3.00003-7
- Smýkal, P., G. Kenicer, A.J. Flavell, J. Corander, O. Kosterin, and R.J. Redden. 2011. Phylogeny, phylogeography and genetic diversity of the *Pisum* genus. *Plant Genet. Res.* 9, 4-18. Doi: 10.1017/S147926211000033X
- Tiemerman, G., A. Mills, C. Whitfield, T. Frew, R. Butler, and S. Murray. 2005. Linkage mapping of QTLs for seed yield, yield components and developmental traits in pea (*Pisum sativum* L.). In: 4th International Crop Science Congress 45(4), 1336-1344. Doi: 10.2135/cropsci2004.0436
- Tiwari, G. and G.R. Lavanya. 2012. Genetic variability, character association and component analysis in F4 generation of fieldpea (*Pisum sativum* var. *arvense* L.). *Karn. J. Agric. Sci.* 25, 173-175.
- Trněný, O., J. Brus, I. Hradilová, A. Rathore, R. Roma, and P. Kopecký. 2018. Molecular evidence for two domestication events in the pea crop. *Genes* 9(11), 535. Doi: 10.3390/genes9110535
- UPOV, Union Internationale pour la Protection des Obtentions Vegetales. 2003. Protocol for distinctness, uniformity and stability tests: peas (*Pisum sativum* L. sensu lato). CPVO-TP/007/1 Final. European Union, Genova.
- Warkentin, T., P. Smýkal, C. Coyne, N. Weeswn, C. Domoney, D.-J. Bing, A. Leonforte, Z. Xuxiao, G.P. Dixit, L. Boros, K.E. McPhee, R.J. McGee, J. Burstin, and T. Ellis. 2015. Pea. In: De Ron, A. (ed.) Grain legumes. Handbook of plant breeding. Vol. 10. Springer, New York, NY. Doi: 10.1007/978-1-4939-2797-5_2
- Wang, F., J. Fu, L. Dong, and Y. Zhu. 2003. Tendril inheritance in semi-leafless pea and its utilization in breeding. *Yi Chuan Hereditas* 25(2), 185-188.
- Wu, X., N. Li, J. Hao, J. Hu, X. Zhang, and M. Blair. 2017. Genetic diversity of chinese and global pea (*Pisum sativum* L.) collections. *Crop Sci.* 57, 1574-1584. Doi: 10.2135/cropsci2016.04.0271