Antibacterial activity of seed extracts of various species of the Annonaceae family cultivated in Colombia

Actividad antibacteriana de extractos de semillas de diversas especies de la familia Annonaceae cultivadas en Colombia

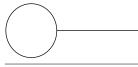


GLORIA EDITH GUERRERO-ÁLVAREZ¹² ANA ISABEL GIRALDO-RIVERA²

Annonaceas fruit species: A) *A. muricata*, B) *A. reticulata*, C) *A. cherimola* and D) *A. montana*. Photo: A.-I. Giraldo-Rivera

ABSTRACT

Public health concerns increase due to microbial propagation and the resistance to existing drugs; therefore, new alternatives are sought, such as the use of natural sources that are antimicrobial agents. Thus, the aim of this study was to evaluate the antibacterial potential of ethanolic extracts of seeds of the species Annona muricata (L.), Annona cherimola (Miller), Annona glabra (L.), Annona reticulata (L.), Rollinia mucosa ([Jacq.] Baillon) and Annona montana (Macfad.) of the Annonaceae family cultivated in Colombia. The bacterial strains correspond to: Staphylococcus aureus (Rosenbach), Enterococcus faecalis ([Andrewes and Horder] Schleifer and Kilpper-Bölz), Bacillus subtilis ([Ehrenberg] Cohn), Escherichia coli ([Migula] Castellani and Chalmers) and Pseudomonas aeruginosa ([J.Schröter] Migula). The antibacterial activity evaluation was performed using the agar diffusion method, each microorganism was inoculated in the medium in a concentration range of 0.2-0.5% v/v and the activity was measured by measuring the inhibition halo. A phytochemical assay was performed to identify the main metabolites to which the activity was attributed. Among the results obtained, it was found the extract of A. montana showed activity against the five bacterial strains, followed by A. glabra; the highest percentage of inhibition achieved was of A. cherimola with 79.86±3.81% activity against E. faecalis; A. reticulata and R. mucosa presented susceptibility only against two bacterial strains, the contrary, the extract of A. muricata did not present any response. The main types of metabolites identified and those attributed to antibacterial potential corresponded to alkaloids, saponins and terpenoids. The above indicates that the plants extract of Annonaceae showed antimicrobial effects. This suggest that species of this family are potential sources of compounds with antibacterial properties, which broadens the knowledge of natural extracts of Colombian materials for use in pharmacotherapy and as alternative for synthetic antibacterial agents.



Additional key words: Annona montana; Annona glabra; bactericide; bacteriostatic; plant extract; inhibition; *Rollinia*.

¹ Universidad Tecnológica de Pereira, Pereira (Colombia). ORCID Guerrero-Álvarez, G.E.: https://orcid.org/0000-0002-0529-5835; Giraldo-Rivera, A.I.: https://orcid.org/0000-0002-4733-5460

² Corresponding author. gguerrero@utp.edu.co



RESUMEN

Los problemas de salud pública aumentan debido a la propagación microbiana y la resistencia a los medicamentos existentes; por lo que se buscan nuevas alternativas, como el uso de fuentes naturales que sean agentes antimicrobianos. Así, el objetivo de este estudio fue evaluar el potencial antibacteriano de extractos etanólico de semillas de las especies Annona muricata (L.), Annona cherimola (Miller), Annona glabra (L.), Annona reticulata (L.), Rollinia mucosa ([Jacq.] Baillon) y Annona montana (Macfad.) de la familia Annonaceae cultivadas en Colombia. Las cepas bacterianas corresponden a: Staphylococcus aureus (Rosenbach), Enterococcus faecalis ([Andrewes and Horder] Schleifer and Kilpper-Bölz), Bacillus subtilis ([Ehrenberg] Cohn), Escherichia coli ([Migula] Castellani and Chalmers) y Pseudomonas aeruginosa ([J.Schröter] Migula). La evaluación de la actividad antibacteriana se realizó por el método de difusión en agar, cada microorganismo se inoculó en el medio en un rango de concentración de 0.2-0.5%, v/v, y se midió la actividad midiendo el halo de inhibición. Se realizó una marcha fitoquímica para identificar los principales metabolitos a los que se atribuyó la actividad. Entre los resultados obtenidos, se encontró que el extracto de A. montana mostró actividad contra las cinco cepas bacterianas, seguido de A. glabra, el mayor porcentaje de inhibición alcanzado fue el de A. cherimola con 79,86±3,81% de actividad frente a E. faecalis; A. reticulata y R. mucosa presentaron susceptibilidad solo frente a dos cepas bacterianas, por el contrario, el extracto de A. muricata no presentó ninguna respuesta. Los principales tipos de metabolitos identificados y a los que se atribuye potencial antibacteriano correspondieron a alcaloides, saponinas y terpenoides. Lo anterior indica que los extractos vegetales de Annonaceae presentaron efectos antimicrobianos. Esto sugiere que las especies de esta familia son fuentes potenciales de compuestos con propiedades antibacterianas, lo que amplía el conocimiento de extractos naturales de materiales colombianos para su uso en farmacoterapia y como alternativa a los agentes antibacterianos sintéticos.

Palabras clave adicionales: *Annona montana; Annona glabra;* bactericida; bacteriostático; extracto vegetal; inhibición; *Rollinia*.

Received: 28-02-2023 Accepted: 23-04-2023 Published: 30-04-2023

$\left(\right)$

Worldwide, one of the greatest public health concerns is the increase in the appearance and spread of microbial pathogens (Malik *et al.*, 2017); in developed countries, half of deaths are due to infectious diseases for which treatment with existing drugs is not enough because of common pathogen resistance (Rodríguez *et al.*, 2017). In the food industry, there have been high degrees of toxicity and carcinogenicity reported for synthetic additives used for food preservation (Khalil *et al.*, 2018).

This problem has a high impact on life, in addition to the cost-benefit of the treatments to treat this pathology, which affects all countries (García-Perez *et al.*, 2021). According to a report by the National Institute of Health (Colombia Instituto Nacional de Salud, 2022), in Colombia, there is an annual increase of 33% in food-borne disease outbreaks compared to the previous years; where diarrheal diseases are the ones with the highest incidence transmitted by microorganisms such as *Salmonella* sp., *Campylobacter*, and *Escherichia coli* ([Migula] Castellani and Chalmers).

INTRODUCTION

This has led to the search for new alternatives, where bioprospecting and the use of natural resources are fundamental tools. Plants have the ability to produce a variety of secondary metabolites with significant antibacterial activity that could break the resistance generated by synthetic antibiotics and efficient in the treatment of bacterial infections (Torres-Chati *et al.*, 2017; Palacios *et al.*, 2019; Mosquera *et al.*, 2020).

In recent years, studies of the Annonaceae family have intensified because are sources of important phytochemical agents with broad potential (Krinski*et al.*, 2014; Giraldo-Rivera and Guerrero-Alvarez, 2019; León-Méndez *et al.*, 2020). Annonaceae is a family of lowland trees that grow mainly in forest and lianas and is found mainly at elevations below 2,000 m. This family has 130 genera, and it is estimated that there are about of 2,450 species in the world, distributed throughout the tropical zones of America, Africa, Indochina and Malaysia; among them there are numerous fruit trees, especially in the genera *Annona* and *Rollinia*. It has four genera of economic interest



in fruit growing: *Annona, Rollinia, Uvaria* and *Asimina* (Maas *et al.*, 2019).

The genus *Annona* is the second-largest genus of Annonaceae in the Neotropics with a total number of 160 species and the most used are *Annona cherimola* (Miller) (chirimoya) and *Annona muricata* (L.) (soursop) for their edible fruits. The importance of the *A. muricata* species in different applications is highlighted. Various parts of this plant are traditionally used to treat cancer, diabetes, hypertension, fever, gastrointestinal disorders, and parasitic infections, as well as the antimicrobial properties of the leaves used to treat viral infections (Pinto *et al.*, 2017).

Among the main metabolites of this family, are acetogenins which have cytotoxic, antitumor, and insecticidal activity (Osorio *et al.*, 2007); isoquinoline alkaloids, which have neurotoxic and antidepressant effects; and, to a lesser extent, phenolic compounds, cyclopeptides, vitamins, glycosides and volatile compounds (Pinto *et al.*, 2017).

In Colombia, the main Annonaceae that are cultivated for its production are: *A. muricata, A. squamosa* and *A. cherimola*. Another minor species found in the wild is *Rollinia mucosa* ([*Jacq.*] *Baillon*). The greatest diversity of species is found in the Amazon (54%), Pacific (27.5%) and Andean (27%) regions. It 87% grow at altitudes of less than 500 m. The Annonaceae family is important from the nutritional point of view, around 4,626.4 ha planted belonging to the Annonaceae family, where 1,700 ha belong to crops of *A. muricata*, located mainly in the departments of Tolima, Valle del Cauca and the Eje Cafetero (León-Méndezet al., 2020).

Therefore, this research was carried out with the purpose of expanding the search for natural sources with antibacterial potential, through the bioprospecting study of Annonacea species cultivated in Colombia. The activity of extract from the seeds of *A. muricata, A. cherimola, Annona glabra, Annona reticulata, R. mucosa* and *Annona montana* was evaluated against the pathogens *S. aureus, E. faecalis, B. subtilis, E. coli* and *P. aeruginosa,* by the agar diffusion method. Additionally, a preliminary phytochemical study was made to establish to which type of compounds the activity found could be attributed.

MATERIALS AND METHODS

Plant material

The seeds were obtained from different Annonaceae ripe fruit species and were collected in different locations in Colombia as indicated in table 1. The taxonomic identification of *A. montana* and *A. glabra* took place in the herbarium at Universidad del Quindío with voucher number 38331 and 38332, respectively. *R. mucosa* was confirmed at the Herbarium of the Universidad de la Amazonia (HUAZ) with voucher number 1525. *A. reticulata* it was identified by Francisco Javier Roldan Palacio, curator of the Herbarium of the University of Antioquia (HUA). *A. cherimola* it was verified by the agronomist Hernán Giraldo of the Santa Rosa de Cabal University Corporation (UNISARC) and *A. muricata* it was verified by the agronomist Juan Felipe Romero.

Scientific name	Common name	Collection site	Latitude - Longitude
Annona muricata (L.)	Soursop	Pereira, Risaralda	4°48′27.3″ N 75°46′0.3″ W
Annona montana (Macfad.)	Mountain soursop	Caicedonia, Valle del Cauca	4°21′11.6″ N 75°51′26″ W
Annona glabra (L.)	Pond apple	Toro, Valle del Cauca	4°41′11.77″ N 76°2′42.05″ W
Rollinia mucosa ([Jacq.] Baillon)	Wild sugar apple	Santo Domingo, Caqueta	1°35′22″ N 75°38′0.2″ W
Annona cherimola (Miller)	Cherimoya	La bella, Risaralda	4°45′55.9″ N 75°38'17″ W
Annona reticulata (L.)	Custard apple	Zarzal, Valle del Cauca	4°23′30.2″ N 76°04′37.3″ W

 Table 1.
 Names of the fruit species of the Annonaceae family cultivated in Colombia.

Obtaining the extracts

Seeds were washed with TEGO 51 (Quios, Bogota) and dried at 37°C for 72 h accordance with that used in previous studies (Giraldo *et al.*, 2020). Then, seeds were crushed in a mill MF 10 Basic (IKA, Campinas, Brazil). The powder obtained was carried out in an extraction process using the passive maceration method at room temperature. It was used absolute ethanol as solvent extraction with a solvent: sample ratio of 1:4 and regular agitation. The extract was filtered and concentrated by rotary evaporator until a viscous consistency and residual solvent was removed by nitrogen flow (Giraldo and Guerrero, 2018).

Antibacterial activity

The antibacterial activity of the Annonaceae extracts was tested against Gram-positive bacteria *Staphylococcus aureus* (Rosenbach) (ATCC 6538), *Enterococcus faecalis* ([Andrewes and Horder] Schleifer and Kilpper-Bölz) (ATCC 51299), and *Bacillus subtilis* ([Ehrenberg] Cohn) (ATCC 21556) and Gram-negative bacteria *Escherichia coli* ([Migula] Castellani and Chalmers) (ATCC 9637) and *Pseudomonas aeruginosa* ([J.Schröter] Migula) (ATCC 9027).

Agar diffusion method (zone inhibition assay). The determination of antibacterial activity was based according to the protocol provided in the Institute of Clinical and Laboratory Standards (CLSI, 2018) and with the methodology by Boorn *et al.* (2010).

Inoculum preparation. The inoculum was prepared from an actively growing culture plate ($Difco^{TM}$ nutrient agar REF. 213000) for 24 h incubate at 37°C; with the aid of a bacteriological loop, colonies were taken and dissolved in distilled water, adjusting the inoculum to a turbidity of 0,5 MacFarland in a DensiCHEK TM Plus.

Inoculation of the culture medium. Agar plates of Mueller-Hinton agar (OXOID, CM0337) were inoculated using a swab from a suspension of each organism containing $1\cdot10^6$ CFU/mL by deep culture technique using Mueller-Hinton agar in 10 cm diameter Petri dishes was implemented, each microorganism was in a concentration at 0.2% v/v by exception from *E. coli* which was prepared at 0.5% v/v. A 3-mm diameter well was cut into the agar once it solidified.

Solutions preparation. The extracts crude was diluted in a small amount of dimethyl sulfoxide (DMSO). Commercial antibiotics were used as positive control. *E. coli* was tested at a concentration of 100 μ g mL⁻¹ amoxicillin (Genfar[®] 2007M-007513-R2); *S. aureus* and *E. faecalis* were tested at a concentration of 200 and 700 μ g mL⁻¹ ampicillin (Genfar[®] 2007M-07219-R2); *P. aeruginosa* and *B. subtilis* were tested at a concentration of 50 and 1000 μ g mL⁻¹ ciprofloxacin (Genfar[®] 2016M-001483-R2). Two controls were employed, a blank containing the same amount of dimethyl sulfoxide (DMSO) was used to dissolve the extracts and ethanol absolute was used a negative control.

Determination antibacterial activity of the extracts. 60 μ L of each extract and the different control to be evaluated was aliquoted into each well. The dishes were incubated at 37°C for 24 h. After incubation, the diameter of the zone of inhibition was measured and the activity was determined by establishing the % inhibition by compared to the reference control. Each sample was tested a minimum of three times and means were calculated.

Phytochemical analysis of Annonaceae extracts

A phytochemical assay was performed, and the analysis of saponins and tannins, reducing sugars, total sugars, flavonoids, and glycosides (Elisha *et al.*, 2017) was performed in tube tests. The analysis of isoquinoline alkaloids, phenols, terpenoids, lignans, sesquiterpene lactones, sterols, anthraquinones, and anthrones was conducted using thin layer chromatography (TLC), as well as the analysis of acetogenins and alkaloids (Giraldo and Guerrero, 2018).

Data analysis

The zone of inhibition (IZD) was measured in millimeters (mm) using the Image J program and the results were expressed as percentage inhibition (%). The inhibition halo of the controls was used as a reference parameter. All experiments were performed in triplicate and data are presented as mean values \pm standard deviation. The statistical differences between the treatments and the control were evaluated using the ANOVA test followed by Bonferroni (P>0.05).

RESULTS AND DISCUSSION

The average yield of seed ethanol extracts the different species of Annonaceae family are indicated in table 2. The range of ethanolic extraction by passive maceration are located between 14 a 40 %.

Species	Seed mass (g)	Extract mass (g)	Yield (%)		
A. muricata	50.0817	7.3412	14.65		
A. montana	31.5852	6.0830	19.26		
A. glabra	18.4550	4.1816	22.65		
R. mucosa	30.2240	6.5534	21.68		
A. cherimola	30.0030	12.0234	40.07		
A. reticulata	30.0047	8.4031	28.01		

of Annonaceae family.

Yield of seed ethanol extracts the different species

Table 2.

The antibacterial activity for six plant extracts against the five bacterial strains examined were assessed by the presence or absence of inhibition zones. The percentage inhibition of species is given in the figure 1. These results indicate that there is no homogenous antimicrobial activity behavior of the plant extracts. The inhibitory activity varied between 10 and 80%. The most sensitive bacterial strain was *B. subtilis,* in which the behavior was different against each extract, with significant differences of the extracts of *A. glabra* and *A. reticulata* that obtained inhibition percentages higher (Fig. 1E), in some cases, no colony growth was observed in the medium, and in others,

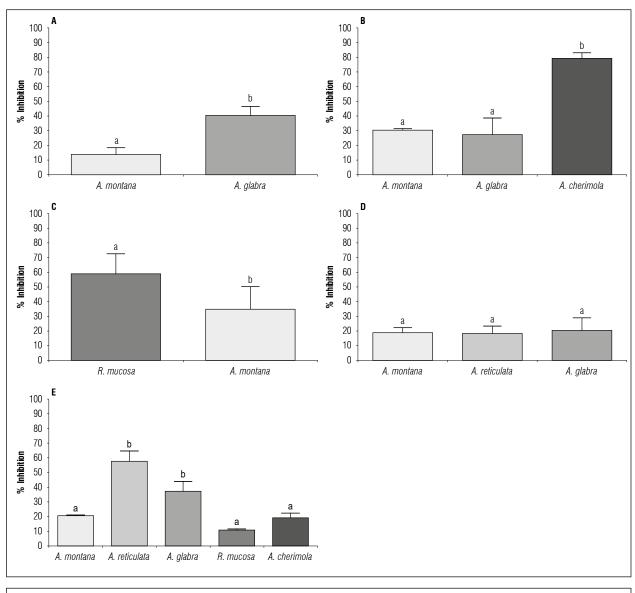


Figure 1. Percentage of inhibition of extract Annonacea against: A) P. aeruginosa, B) E. faecalis, C) S. aureus, D) E. coli, E) B. subtilis. Different letters indicate significant differences between extracts according to Bonferroni (P≤0.05), n=3±SE. translucent dotted halos were observed (Fig. 2). On the other hand, *P. aeruginosa* and *E. coli* were the most resistant to the extracts analyzed, however, percentage inhibition of *A. glabra* of 40% was observed in *P. aeruginosa* (Fig. 1A).

The *A. montana* extract presented activity against the five bacterial strains, where the most important results were against *S. aureus* ($34.87 \pm 15.37\%$) and *E. faecalis* ($30.61 \pm 1.04\%$). The *A. cherimola* extract exhibited the highest percentage of inhibition against *E. faecalis* ($79.86 \pm 3.81\%$), with significant differences with the results obtained by the extracts of the other two species (*A. montana* with $30.60 \pm 1.04\%$ and *A. glabra* with 27.75 \pm 6.69\%) (Fig. 1B).

The extracts of the species *A. montana, A. reticulata* and *A. glabra* were the only ones that showed activity against *S. aureus*, without presenting significant differences between the percentages of inhibition. On the other hand, the extract of *A. muricata* was the only one that did not present any activity against the microorganisms evaluated in this study.

The chemical analysis of the Annonaceae crude extracts showed the presence of isoquinoline alkaloids, anthrones, glycosides, and acetogenins for the six species tested. Only *A. montana* and *A. glabra* were positive for sesquiterpene lactones but negative for sterols. Sugars were present in *A. muricata, A. glabra* and *A. montana*, and saponins were detected only in *A. cherimoya, A. reticulata* and *A. montana*. This allowed the detection of the presence of different phytochemical constituents (Tab. 2) that could be responsible for the antibacterial activity.

In general, it was observed that the Annonaceae extracts evaluated had higher activity against Grampositive bacterial strains. This may be because Gram-negative strains are more resistant due to the structure of the cell wall, which acts as a permeability barrier, reducing the uptake of compounds into the cell (Elisha et al., 2017). One of the most important results found in this study is the activity of A. cherimola against E. faecalis (79.86±3.81 %) and that of R. mucosa against E. coli (58.95±13.63). It is important to highlight the result obtained by *A. reticulata* against *B.* subtilis (Fig. 2), where a halo of greater growth of the microorganism was observed around the well, a phenomenon that should be studied in greater depth. A. cherimola is a fruit known for its high cytotoxic potential due to the presence of acetogenins, compounds that are characterized by the activation of apoptotic pathways, which leads to cell death (Haykal et al.,

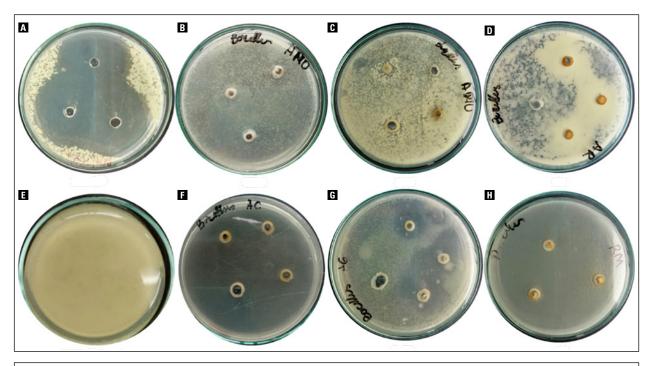


Figure 2. Inhibition of the growth of *B. subtilis* against (A) Antibiotic (B) *A. montana* (C) *A. muricata* (D) *A. reticulata* (E) Growth control (F) *A. cherimola* (G) *A. glabra* (H) *R. mucosa*.



Metabolites	A. muricata	R. mucosa	A. cherimola	A. glabra	A. reticulata	A. montana
Isoquinoline alkaloids	+	+	+	+	+	+
Phenols and flavonoids	-	-	-	-	-	-
Terpenoids and lignans	+	+	-	+	+	+
Sesquiterpene lactones	-	-	-	+	-	+
Sterols	+	+	+	-	+	-
Terpenoids	-	-	+	-	-	-
Anthraquinones	-	-	-	-	-	-
Anthrones	+	+	+	+	+	+
Coumarins	-	-	-	-	-	-
Acetogenins	+	+	+	+	+	+
Alkaloids	+	+	+	+	+	+
Reducing sugars	+	-	-	-	-	-
Total sugars	+	-	-	+	-	+
Flavonoids	-	-	-	-	-	-
Tannins	-	-	-	-	-	-
Glycosides	+	+	+	+	+	+
Saponins	-	-	+	-	+	+

Tahle 2	Phytochemical analysis of extracts from Annonaceae specie	2
1auic 2.	"IIYLUCIIGIIIICAI AIIAIYSIS UI GALIACLS IIVIII AIIIIVIIACGAG SPECIE	·ə.

2019). This allows establishing these compounds as responsible for the activity presented by these fruits, as has been reported in other studies, the acetogenesis of this fruit presents an important activity against gram positive bacteria (Aguilar-Villalva *et al.*, 2021; Perrone *et al.*, 2022).

R. mucosa is considered an exotic Amazonian fruit. Previous reports have indicated that the hexane extracts of the seeds, pulp and stems were evaluated against five microorganisms, where the highest result corresponded to stem extracts against *Candida albicans* with a percentage of inhibition of 39.14% (Fernández *et al.*, 2020), being the result obtained in this study promising for this fruit species.

On the other hand, the extracts of the *A. reticulata* leaf have shown inhibitory activity against *P. aeruginosa*, identifying compounds of the alkaloid, flavonoid, phenol, steroid and tri-terpene type (Harahap *et al.*, 2022), also identify in this study. Terpenes are attributed to the disruption capacity of the bacterial cell membrane (Saleem *et al.*, 2003). Saponins have been associated with the ability to produce pores in the cell membrane, forming complexes with cholesterol, altering cell morphology, especially in Grampositive bacteria (Arabski *et al.*, 2009). These two

types of compounds could explain the result observed with this extract compared to *B. subtilis.*

It is important to comment that A. muricata has been the most studied species, where the polar extracts of the leaves with the presence mainly of isoquinoline alkaloids act against multiresistant bacteria, especially E. faecalis, E. coli and P. aeruginosa with promising results (Tojola et al., 2019; Iyanda-Joel et al., 2019; Aguilar-Hernández et al., 2023). Although, in this study, A. muricata did not present bactericidal activity, this can be attributed to the fact that the extract evaluated was from the seed, with other types of metabolites present. Another example of this is A. glabra where stem bark extracts, flavonoid-like compounds with antimicrobial properties against P. aeruginosa have been identified (Galvão et al., 2016) and this allows highlighting the importance of the type of extract obtained and the type of functional groups identified in the preliminary phytochemical analysis.

Additionally, it is observed that the response of the microorganisms against the extracts was diverse, especially with *E. coli* and *B. subtilis*. The growth of the microorganisms presented visual changes or in some cases it was inhibited, indicating that the

antimicrobial activity of the extracts could be associated with the effect on the target site on the cell walls as well as with the type of interacting metabolites. The mechanisms include the inhibition of the cell wall, the blocking resistance mechanisms, the inhibition of protein synthesis, and metabolism (Calvo and Martínez-Martínez, 2009).

The results obtained in the phytochemical assays suggest that the Annonaceae extracts may have different antimicrobial activities because differences in the composition of the secondary metabolites were found. According to previous studies, the antimicrobial activity depends on the type of metabolite present (Maillard, 2002), which can exhibit different modes of action or be associated with microorganisms that more effectively overcome the effect of a compound or adapt to it (Berić *et al.*, 2018; Çördük *et al.*, 2017; Gavrilović *et al.*, 2016; Ji *et al.*, 2016; Maya *et al.*, 2021).

All the evaluated extracts tested positive for alkaloids and glycosides; alkaloids are good bactericidal agents and exert their activity on the membrane. These compounds disrupt the formation of the bacterial Z-ring and inhibit bacterial cytokinesis, which ultimately leads to inhibition of cell reproduction (Barbieri et al., 2017; Nájera-Arce et al., 2018). Additionally, different mechanisms of action have been proposed for glycosides, with indications that they act mainly in the membrane to alter cellular permeability and as chelating agents and inhibit the production of toxins and microbial growth (Mullins, 1990; Rabea et al., 2003). Sterol-derived compounds have been reported as bacteriostatic agents because they have strong inhibitory effects (Hovenkamp et al., 2008; Bhardwaj et al., 2014).

Although not all extracts showed activity against the different strains evaluated in this study, it has been reported that extracts from species of the Annonaceae family can modulate drug resistance (Barboza *et al.*, 2015). These results indicate that the extracts have good potential for therapeutic uses against some pathogens.

CONCLUSION

According to the results, ethanolic extracts obtained from the seeds of the main Colombian species of Annonaceae showed antimicrobial properties against the microorganisms *S. aureus*, *E. faecalis*, *B. subtilis*, *E. coli* and *P. aeruginosa*; it suggests that different extracts contain compounds with antibacterial properties, and it is according to their phytochemical differences. These results contribute to the bioprospecting of this family since previous studies have been carried out mainly with extracts obtained from the leaves.

Owing to the potential antibacterial actions of Annonaceae fruit, it is possible to continue for further studies aimed identifying the single active principles and evaluating possible synergism of antimicrobial activity among these extracts. The applications could an accessible and safe alternative to synthetic antimicrobial drugs.

Conflict of interests: The manuscript was prepared and reviewed with the participation of the authors, who declare that there exists no conflict of interest that puts at risk the validity of the presented results.

BIBLIOGRAPHIC REFERENCES

- Aguilar-Hernández, G., B.A. López-Romero, A. Pérez-Larios, J.M. Ruvalcaba-Gómez, I. Castellanos-Huerta, G. Tellez-Isaias, V.M. Petrone-García, L.M. Anaya-Esparza, and E. Montalvo-González. 2023. Antibacterial activity of crude extract and purified acetogenins from *Annona muricata* seeds. Appl. Sci. 13(1), 558. Doi: https://doi.org/10.3390/app13010558
- Aguilar-Villalva, R., G.A. Molina, B.L. España-Sánchez, L.F. Díaz-Peña, A. Elizalde-Mata, E. Valerio, C. Aranza-Ricardo, and M. Estevez. 2021. Antioxidant capacity and antibacterial activity from *Annona cherimola* phytochemicals by ultrasound-assisted extraction and its comparison to conventional methods. Arab. J. Chem. 14(7), 103239. https://doi.org/10.1016/j. arabjc.2021.103239
- Arabski, M., S. Wąsik, K. Dworecki, and W. Kaca. 2009. Laser interferometric and cultivation methods for measurement of colistin/ampicilin and saponin interactions with smooth and rough of *Proteus mirabilis* lipopolysaccharides and cells. J. Microbiol. Methods 77(2), 178-183. Doi: https://doi.org/10.1016/J. MIMET.2009.01.020
- Barbieri, R., E. Coppo, A. Marchese, M. Daglia, E. Sobarzo-Sánchez, S.F. Nabavi, and S.M. Nabavi. 2017. Phytochemicals for human disease: An update on plant-derived compounds antibacterial activity. Microbiol. Res. 196, 44-68. Doi: https://doi.org/10.1016/J. MICRES.2016.12.003
- Barboza, T.J.S., A.F. Ferreira, A.C.P.R. Ignacio, and N. Albarello. 2015. Antimicrobial activity of Anonna mucosa (Jacq.) grown in vivo and obtained by in vitro culture. Braz. J. Microbiol. 46(3), 785-789. Doi: https://doi. org/10.1590/S1517-838246320140468

- Berić, T., M. Biočanin, S. Stanković, I. Dimkić, T. Janakiev, D. Fira, and J. Lozo. 2018. Identification and antibiotic resistance of *Bacillus* spp. isolates from natural samples. Arch. Biol. Sci. 70(3), 581-588. Doi: https://doi. org/10.2298/ABS180302019B
- Bhardwaj, R., A. Yadav, P. Sharma, and R.A. Sharma. 2014. In vitro and in vivo GC-MS profile and antimicrobial activity of phytosterols of Datura stramonium. Res. J. Med. Plants 8(3), 112-120. Doi: https://doi. org/10.3923/rjmp.2014.112.120
- Boorn, K.L., Y.-Y. Khor, E. Sweetman, F. Tan, T.A. Heard, and K.A. Hammer. 2010. Antimicrobial activity of honey from the stingless bee *Trigona carbonaria* determined by agar diffusion, agar dilution, broth microdilution and time-kill methodology. J. Appl. Microbiol. 108(5), 1534-1543. Doi: https://doi. org/10.1111/j.1365-2672.2009.04552.x
- Calvo, J. and L. Martínez-Martínez. 2009. Mecanismos de acción de los antimicrobianos. Enferm. Infecc. Microbiol. Clin. 27(1), 44-52. https://doi.org/10.1016/j. eimc.2008.11.001
- Colombia Instituto Nacional de Salud. 2022. Protocolo de vigilancia de brotes de enfermedades transmitidas por alimentos, versión 4. Doi: https://doi.org/10.33610/ infoeventos.14
- CLSI, Clinical and Laboratory Standards Institute. 2018. Performances standards for antimicrobial disk susceptibility test. 13th ed. CLSI document M02-A12. Wayne, PA.
- Çördük, N., S. Demirbas, and D.N. Hacioglu. 2017. A comparative study of the antimicrobial properties and antioxidant enzyme activities of field-grown and *in vitro*-propagated plants of endemicdigitalis trojana Ivanina. Arch. Biol. Sci. 69(4), 603-610. Doi: https:// doi.org/10.2298/ABS161216004C
- Elisha, I.L., F.S. Botha, L.J. McGaw, and J.N. Eloff. 2017. The antibacterial activity of extracts of nine plant species with good activity against *Escherichia coli* against five other bacteria and cytotoxicity of extracts. BMC Complement. Altern. Med. 17(1), 133. Doi: https:// doi.org/10.1186/s12906-017-1645-z
- Fernández, I.M., E.A. Chagas, S.A.S. Maldonado, J.A. Takahashi, R.S. Alemán, A.A. Melo Filho, R.C. Santos, P.R.E. Ribeiro, J.A.M. Fuentes, P.C. Chagas, and A.C.G. R. Melo. 2020. Antimicrobial activity and acetilcolinesterase inhibition of oils and Amazon fruit extracts. J. Med. Plants Res. 14(3), 88-97. https://doi. org/10.5897/JMPR2019.6790
- Galvão, S.S.L., A.S. Monteiro, E.P. Siqueira, M.R.O. Bomfim, M.V. Dias-Souza, G.F. Ferreira, A.M.L. Denadai, Á.R.C. Santos, V.L. Santos, E.M. Souza-Fagundes, E.S. Fernandes and V. Monteiro-Neto. 2016. Annona glabra flavonoids act as antimicrobials by binding to Pseudomonas aeruginosa cell walls. Front. Microbiol. 7, 2053. Doi: https://doi.org/10.3389/fmicb.2016.02053

- García-Perez, J., J.B. Ulloa-rojas, and S. Mendoza-Elvira. 2021. Patógenos bacterianos y su resistencia a los antimicrobianos en los cultivos de tilapia en Guatemala. Uniciencia 35(2), 1-17. Doi: https://doi.org/10.15359/ ru.35-2.4
- Gavrilović, M., M.D. Soković, M. Stanković, P.D. Marin, Z. Dajić-Stevanović, and P. Janaćković. 2016. Antimicrobial and antioxidative activity of various leaf extracts of *Amphoricarposvis*. (Asteraceae) taxa. Arch. Biol. Sci. 68(4), 803-810. Doi: https://doi.org/10.2298/ ABS160112068G
- Giraldo, A.I. and G.E. Guerrero. 2018. Rollinia mucosa (Jacq.) Baillon (Annonaceae) active metabolites as alternative biocontrol agents against the lace bug Corythucha gossypii (Fabricius): An insect pest. Univ. Sci. 23(1), 21-34. Doi: https://doi.org/10.11144/Javeriana.SC23-1.rmjb
- Giraldo, A.I., G.E. Guerrero, J.P. Arrubla, L.M. Baena, D. Paredes, and M.A. Gomez. 2020. The effects of Annonaceae and Amaryllidaceae extracts in controlling the Thrips tabaci Lindeman (Thysanoptera: Thripidae). Rev. Bras. Cienc. Agrar. 15(2), 1-9. Doi: https://doi. org/10.5039/agraria.v15i2a6933
- Giraldo-Rivera, A.I. and G.E. Guerrero-Alvarez. 2019. Botanical biopesticides: Research and development trends, a focus on the *Annonaceae* family. Rev. Colomb. Cienc. Hortic. 13(3), 371-383. Doi: https://doi.org/10.17584/ rcch.2019v13i3.9489
- Harahap, D., S. Niaci, V. Mardina, B. Zaura, I. Qanita, A. Purnama, K. Puspita, D.R. Rizki, and M. Iqhrammullah. 2022. Antibacterial activities of seven ethnomedicinal plants from family *Annonaceae*. J. Adv. Pharm. Technol. Res. 13(3), 148-153.
- Haykal, T., P. Nasr, M.H. Hodroj, R.I. Taleb, R. Sarkis, M.N.E. Moujabber, and S. Rizk. 2019. Annona cherimola seed extract activates extrinsic and intrinsic apoptotic pathways in leukemic cells. Toxins 11(9), 506. Doi: https://doi.org/10.3390/toxins11090506
- Hovenkamp, E., I. Demonty, J. Plat, D. Lütjohann, R.P. Mensink, and E.A. Trautwein. 2008. Biological effects of oxidized phytosterols: A review of the current knowledge. Prog. Lipid Res. 47(1), 37-49. Doi: https:// doi.org/10.1016/j.plipres.2007.10.001
- Iyanda-Joel, W.O., E.A. Omonigbehin, E.E.J. Iweala, and S.N. Chinedu. 2019. Antibacterial studies on fruitskin and leaf extracts of *Annona muricata* in Ota, Nigeria. IOP Conf. Ser.: Earth Environ. Sci. 331(1), 120-129. https://doi.org/10.1088/1755-1315/331/1/012029
- Ji, J., J. Yang, L. Huang, and Z. Kang. 2016. A novel antifungal peptide purified from *Bacillus subtilis* strain EDR4. Arch. Biol. Sci. 68(2), 319-324. Doi: https://doi. org/10.2298/ABS150325022J
- Khalil, N., M. Ashour, S. Fikry, A.N. Singab, and O. Salama. 2018. Chemical composition and antimicrobial activity of the essential oils of selected *Apiaceous* fruits.

Future J. Pharm. Sci. 4(1), 88-92. Doi: https://doi.org/10.1016/J.FJPS.2017.10.004

- Krinski, D., A. Massaroli, and M. Machado. 2014. Potencial inseticida de plantas da familia Annonaceae. Rev. Bras. Frutic. 36(Spl. 1), 225-242. Doi: https://doi. org/10.1590/S0100-29452014000500027
- León-Méndez, G., N. Pajaro-Castro, and C. Granados-Conde. 2020. Annona squamosa una fuente de acetogeninas en la actualidad. Ciencia y Salud Virtual 12(2), 88-101.
- Maas, P.J.M., L.Y.T. Westra, L.W. Chatrou, N. Verspagen, H. Rainer, N.A. Zamora, and R.H.J. Erkens. 2019. Twelve new and exciting Annonaceae from the Neotropics. PhytoKeys 126, 25-69. Doi: https://doi.org/10.3897/ phytokeys.126.33913
- Maillard, J.-Y. 2002. Bacterial target sites for biocide action. J. Appl. Microbiol. 92(Spl. 1), 16-27. Doi: https://doi. org/10.1046/j.1365-2672.92.5s1.3.x
- Malik, T.A., A.N. Kamili, M.Z. Chishti, S. Ahad, M.A. Tantry, P.R. Hussain, and R.K. Johri. 2017. Breaking the resistance of *Escherichia coli*: Antimicrobial activity of *Berberis lycium* Royle. Microb. Pathog. 102, 12-20. Doi: https://doi.org/10.1016/j.micpath.2016.11.011
- Maya, C.A., T.A. Madrid, and J.E. Parra. 2021. Efecto de Bacillus subtilis sobre metabolitos sanguíneos y parámetros productivos en pollo de engorde. Biotecnol. Sector Agropecuario Agroind. 19(1), 105-116. Doi: https://doi.org/https://doi.org/10.18684/bsaa.v19. n1.2021.1468
- Mosquera, T., A. Medrano, and M.E. Maldonado. 2020. Extractos naturales una alternativa conservante en la industria cosmética. Rev. Iber. Sist. Tecnol. Inf. (E30), 139-149.
- Mullins, J.T. 1990. Regulatory mechanisms of β-glucan synthases in bacteria, fungi, and plants. Physiol. Plant. 78(2), 309-314. Doi: https://doi.org/10.1111/j.1399-3054.1990.tb02096.x
- Nájera-Arce, C., P. Álvarez-Fitz, D. Pérez-Castro, J. Toribio-Jiménez, and N. Castro-Alarcón. 2018. Actividad antibacteriana de diatomeas marinas aisladas de Acapulco, Guerrero, México. Rev. Biol. Mar. Oceanogr. 53(2), 195-207. Doi: https://doi.org/10.22370/ rbmo.2018.53.2.1293
- Osorio, E., G.J. Arango, N. Jiménez, F. Alzate, G. Ruiz, D. Gutiérrez, M.A. Paco, A. Giménez, and S. Robledo. 2007.

Antiprotozoal and cytotoxic activities *in vitro* of Colombian Annonaceae. J. Ethnopharmacol. 111(3), 630-635. Doi https://doi.org/10.1016/j.jep.2007.01.015

- Palacios, S., R. Rodríguez, A.C. Flores, M.L. Chávez, R. Ramos, E.P. Segura, and A. Ilina. 2019. Alternativas para el control de bacterias transmitidas por alimentos. CienciAcierta 57, 1-10.
- Perrone, A., S. Yousefi, A. Salami, A. Papini, and F. Martinelli. 2022. Botanical, genetic, phytochemical and pharmaceutical aspects of *Annona cherimola* Mill. Sci. Hortic. 296, 110896. Doi: https://doi.org/10.1016/j. scienta.2022.110896
- Pinto, N.C.C., L.M. Campos, A.C.S. Evangelista, A.S.O. Lemos, T.P. Silva, R.C.N. Melo, C.C. Lourenço, M.J. Salvador, A.C.M. Apolônio, E. Scio, and R.L. Fabri. 2017. Antimicrobial *Annona muricata* L. (soursop) extract targets the cell membranes of Gram-positive and Gram-negative bacteria. Ind. Crops Prod. 107, 332-340. Doi: https://doi.org/10.1016/j.indcrop.2017.05.054
- Rabea, E.I., M.E.-T. Badawy, C.V. Stevens, G. Smagghe, and W. Steurbaut. 2003. Chitosan as antimicrobial agent: Applications and mode of action. Biomacromolecules 4(6), 1457-1465. Doi: https://doi.org/10.1021/ bm034130m
- Rodríguez, C.N., A.G. Zarate, and L.C. Sánchez. 2017. Actividad antimicrobiana de cuatro variedades de plantas frente a patógenos de importancia clínica en Colombia. Nova 15(27), 119-129.
- Saleem, M., M. Nazir, M.S. Ali, H. Hussain, Y.S. Lee, N. Riaz, and A. Jabbar. 2003. Antimicrobial natural products: an update on future antibioticdrug candidates. Nat. Prod. Rep. 27(2), 238-254. Doi: https://doi. org/10.1039/B916096E
- Torres-Chati, J., J. León-Quispe, and G. Tomas-Chota. 2017. Actividad antibacteriana y antifúngica de extractos de hojas de *Luma chequen* (Molina) A. Gray arrayán frente a patógenos de origen clínico. Rev. Soc. Ven. Microbiol. 37(1), 10-16.
- Tojola, O.B., L. Lajide, B.J. Owolabi, M. Olaleye, and S.O. Okoh. 2019. Antifungal activities of ethyl acetate and methanol extracts of *Annona muricata* aerial part. Int. J. Innov. Res. Dev. 8(8), 41-44. Doi: https://doi. org/10.24940/ijird/2019/v8/i8/aug19027