

# Evaluation of Activated Carbon Electrodes as Anodes in a Microbial Fuel Cell Using *Shewanella* *Putrefaciens*

Diana-Marcela Vanegas-Hernández; Mónica-Liliana Cardona-  
Aristizabal; Zulamita Zapata-Benabithé

**Citation:** D.-M. Vanegas-Hernández, M.-L. Cardona-Aristizabal, and Z. Zapata-Benabithé, “Evaluation of Activated Carbon Electrodes as Anodes in a Microbial Fuel Cell Using *Shewanella Putrefaciens*,” *Revista Facultad de Ingeniería*, vol. 29 (54), e10468, 2020.  
<https://doi.org/10.19053/01211129.v29.n54.2020.10468>

**Received:** December 22, 2019; **Accepted:** May 2, 2020;  
**Published:** May 13, 2020

**Copyright:** This is an open access article distributed under license [CC](#)

[BY](#)



**Conflict of interest:** The authors state there is no conflict of interest.

# Evaluation of Activated Carbon Electrodes as Anodes in a Microbial Fuel Cell Using *Shewanella Putrefaciens*

Diana-Marcela Vanegas-Hernández<sup>1</sup>

Mónica-Liliana Cardona-Aristizabal<sup>2</sup>

Zulamita Zapata-Benabithé<sup>3</sup>

## Abstract

In this work, three types of activated carbons were evaluated as electrodes in the anode chamber of a two-chamber microbial fuel cell (MFC). The evaluation was applied using a pure *Shewanella Putrefaciens* culture due to its gram-negative characteristics. In the cathode chamber, a platinum electrode was used, and a Nafion® 117 proton exchange membrane was selected as a separator of both chambers. The activated carbons were obtained from different precursors (coffee husk, commercial coal, and mineral coal), with different microporous and surface properties. From the voltage and current measurements, it was found that the cell power values varied between 0.008 mW and 0.045 mW. The electrode obtained from chemical activation of coffee husk with H<sub>3</sub>PO<sub>4</sub> at 450 °C (Q) showed the best electrochemical behaviour and highest power values. This result may be mainly related to the macroscopic morphology and mesopores that improve the wettability of the surface by the medium thought carbonaceous material. SEM images showed a better biofilm formation, larger filaments of the bacteria, and micro-beds formation over the surface of bio-anode Q, which improved the interaction with the

---

<sup>1</sup> Ph. D. Universidad Pontificia Bolivariana (Medellín-Antioquia, Colombia). [diana.vanegas@upb.edu.co](mailto:diana.vanegas@upb.edu.co). ORCID: [0000-0002-9519-0841](https://orcid.org/0000-0002-9519-0841)

<sup>2</sup> M. Sc. Universidad Pontificia Bolivariana (Medellín-Antioquia, Colombia). [monica.cardona@upb.edu.co](mailto:monica.cardona@upb.edu.co). ORCID: [0000-0002-5829-3438](https://orcid.org/0000-0002-5829-3438)

<sup>3</sup> Ph. D. Universidad Pontificia Bolivariana (Medellín-Antioquia, Colombia). [zulamita.zapata@upb.edu.co](mailto:zulamita.zapata@upb.edu.co). ORCID: [0000-0002-4497-4865](https://orcid.org/0000-0002-4497-4865)

microorganism, its metabolism, and electrons extracellular transfer. Therefore, activated carbon from coffee husk could be considered as a promising material for electrodes of microbial fuel cells.

**Keywords:** activated carbon; anodic chamber; carbonaceous materials; mediators microorganism; *Shewanella Putrefaciens*; two chambers cells.

## **Evaluación de electrodos de carbón activado como ánodos de una celda de combustible microbiana usando *Shewanella Putrefaciens***

### **Resumen**

En este trabajo se evaluaron tres tipos de carbones activados como electrodos en la cámara anódica de una celda de combustible microbiana (MFC) de dos cámaras. Se utilizó un cultivo de *Shewanella putrefaciens* puro debido a sus características gran-negativa. En la cámara catódica se utilizó un electrodo de platino y Nafion® 117 como membrana de intercambio de protones para separar las dos cámaras. Los carbones activados seleccionados presentaron diferentes precursores (cisco de café, carbón comercial, y carbón mineral), con diferentes morfologías y propiedades porosas y superficiales. A partir de las mediciones de voltaje y corriente se encontró que los valores de potencia de la celda variaron entre 0.008 mW y 0.045 mW. El electrodo preparado utilizando carbón activado a partir de cisco de café (Q) presentó el mejor comportamiento electroquímico y valores de potencia más altos; este carbón se obtuvo a partir de la activación de cisco de café con H<sub>3</sub>PO<sub>4</sub> a 450 °C. La morfología macroscópica de este carbón activado permitió una mejor migración del medio de cultivo acuoso hacia los macroporos y posteriormente mesoporos que mejoran la mojabilidad de la superficie. Las microfotografías SEM mostraron una mejor formación de la biopelícula, filamentos más largos de la bacteria y la formación de microlechos en el bio-ánodo Q, que favoreció la interacción con *el microorganismo*, facilitando su metabolismo y la transferencia extracelular de electrones. Por tal motivo, el carbón activado a partir de cisco de café podría ser considerado como un material prometedor para su implementación como electrodos en celdas de combustible microbiana.

**Palabras clave:** cámara anódica; carbón activado; celdas dos cámaras; materiales carbonosos; microorganismos mediadores; *Shewanella Putrefaciens*.

### **Avaliação de eletrodos de carvão ativado como ânodos de uma célula de combustível microbiana usando *Shewanella Putrefaciens***

#### **Resumo**

Neste trabalho avaliaram-se três tipos de carvões ativados como eletrodos na câmara anódica de uma célula de combustível microbiana (MFC) de duas câmaras. Utilizou-se um cultivo de *Shewanella putrefaciens* puro devido a suas características gram-negativa. Na câmara catódica utilizou-se um eletrodo de platino e Nafion® 117 como membrana de intercâmbio de prótons para separar as duas câmaras. Os carvões ativados selecionados apresentaram diferentes precursores (cisco de café, carvão comercial, e carvão mineral), com diferentes morfologias e propriedades porosas e superficiais. A partir das medições de voltagem e corrente encontrou-se que os valores de potência da célula variaram entre 0.008 mW e 0.045 mW. O eletrodo preparado utilizando carvão ativado a partir de cisco de café (Q) apresentou o melhor comportamento eletroquímico e valores de potência mais altos; este carvão obteve-se a partir da ativação de cisco de café com H<sub>3</sub>PO<sub>4</sub> a 450 °C. A morfologia macroscópica deste carvão ativado permitiu uma melhor migração do meio de cultivo aquoso em direção aos macroporos e posteriormente mesoporos que melhoram a molhabilidade da superfície. As microfotografias SEM mostraram uma melhor formação da biopelícula, filamentos mais longos da bactéria e a formação de microleitons no bio-ânodo Q, que favoreceu a interação com o *micro-organismo*, facilitando seu metabolismo e a transferência extracelular de elétrons. Por tal motivo, o carvão ativado a partir de cisco de café poderia ser considerado como um material promissor para sua implementação como eletrodos em células de combustível microbiana.

**Palavras chave:** câmara anódica; carvão ativado; células duas câmaras; materiais carbonosos; micro-organismos mediadores; *Shewanella Putrefaciens*.

## I. INTRODUCTION

Renewable energy is increasing and required in order to decrease dependence on fossil fuels, reduce environmental impact and help developing countries to provide more reliable and safer energy systems [1]. Energy consumed in Colombia that comes from renewable sources (common use of biomass and hydroelectric) is around 22 %, and in a lower percentage the energy that comes from the sun, wind, sea, or biomass for conversion into thermal/electric energy [1-2].

Microbial fuel cells (MFCs) are a green technology and a promising solution to meet future energy needs. On a large scale, this type of biotechnology can be incorporated into domestic and industrial wastewater treatment and electricity production from organic wastes, and to bioremediation of contaminated environments at low cost and efficiently [3]. Also, they can operate at room temperature, do not require gas treatment, and have a wide potential in areas not electrically interconnected [4].

The microorganisms in MFCs convert chemical energy to electrical energy, by oxidizing a substrate at the anode as part of their metabolism. Under anaerobic conditions  $\text{CO}_2$ , protons and electrons are produced, and under aerobic conditions only  $\text{CO}_2$  and water are produced [5-6]. Power output from these systems are generally low due to their complex design, high internal resistance, and electrode losses [7].

Chemically stable, biocompatible, and conductive materials are used for the anode in the reactor solution, such as carbon products due to their high stability, electrical conductivity, and surface area. Carbon cloth, carbon or graphite felt, activated carbons, carbon paper, graphite fiber “brush”, carbon granules or spheres have been used as electrodes [8-11]. Some microorganisms have direct electron transfer properties from inside the cell to acceptors via highly conductive type-C cytochromes, biofilms, and nanowires. These microorganisms have high coulombic efficiency and can form biofilms on the anode surface, transferring the electrons directly to the anode and resulting in the production of more energy [12].

This work evaluates the production of electrical energy in a MFC on a laboratory scale. We will be using activated carbons as anodes in a chamber that contains a

conventional medium suitable for *Shewanella putrefaciens*. This microorganism has previously been used as an electrogenic organism [13-15], given its versatility of growth, and anaerobic and aerobic powers. Furthermore, it is a Gram-negative bacterium and can transfer electrons directly to the electrodes to generate small electrical currents.

## II. MATERIALS AND METHODS

The following section describes the anode chamber construction, morphological and superficial characterization of the activated carbons used as anodes, and the electrochemical measurements carried out in the MFC.

### **A. Microbial Strains, Medium and Cultivation**

Bacterial strain *S. putrefaciens* ATCC 8071 was held in cryovials, cultivated aerobically in 50 mL of Luria Bertani (LB) medium (Bacto Tryptone, Yeast Extract, NaCl and supplemented with Lactate at pH 7.4). Bacterial cells were harvested by centrifugation (1500 rpm, 15 min, 4 ° C) and washed with buffer solution, after 36-48 hr of incubation at 160 rpm and 30 °C. Finally, the cells were suspended in 250 ml of LB broth at  $1.5 \times 10^8$  cells / ml.

### **B. Synthesis, Characterization and Surface Morphology of Activated Carbons**

Material Q is an activated carbon (CA) obtained from coffee husk by using phosphoric acid ( $H_3PO_4$ ) at 85% as an activating agent with mass ratio of husk/ $H_3PO_4$  of 2/1 at 500 °C for 1 hr 5 °C/min. Material A is a CA obtained from anthracite, using 6M potassium carbonate ( $K_2CO_3$ ) as an activating agent with mass ratio of Anthracite/ $K_2CO_3$  of 1/1 to 800 °C for 1 h at 5 °C/min. Finally, material P is a commercial CA from Protokimica S.A.S.

Carbonaceous materials were morphologically characterized by scanning electron microscopy (SEM) in a Jeol, JCM-6000plus, porous and superficial from the construction of  $N_2$  isotherms at -196 ° C in an ASAP 2020 Plus (Micromeritics). Specific surface area (ASE) was determined from the isotherm applying the BET model ( $S_{BET}$ ), size ( $D_{micro}$ ) and the volume of micropores ( $V_{micro}$ ) applying Dubinin-

Radushkevich model, the volume of mesopores applying the Gurdovich ( $V_{\text{meso}}$ ). Additionally, bacterial growth on the surface of the carbon electrodes was analyzed by SEM.

### C. Microbial Fuel Cell

Figure 1 shows a microbial fuel cell diagram with two chambers, each one of 250 ml volume, separated by a membrane of 2 cm diameter. Culture medium was introduced into anode chamber with 1.6 g of CA deposited inside a rectangular mesh (2 cm × 3 cm) of stainless steel to allow continuous contact of the medium with the activated carbons.

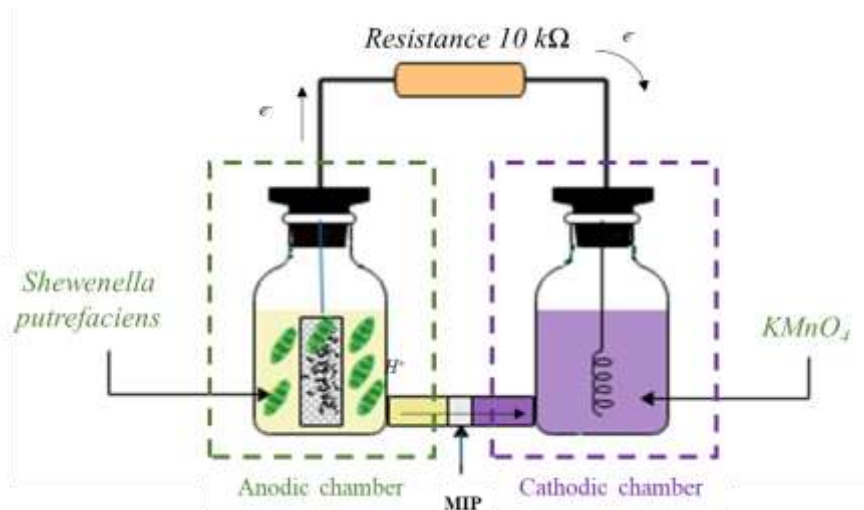


Fig. 1. Two-chamber microbial fuel cell.

Both chambers are separated by a conventional proton exchange membrane (MIP), Nafion® 117 [3]. Membranes were previously activated with 50 ml of a 1% saline solution at 40 °C for 3 hours. In the cathodic chamber, 0.5 mm diameter platinum wire with a length of 25 cm was used in a 0.1 M solution of potassium permanganate ( $\text{KMnO}_4$ ) as an acceptor electrode [16]. The electrons produced in the anode chamber flow through an external resistance of 10 k $\Omega$  before reaching the cathode.

Voltage and current of the electrochemical cell were measured by connecting a multimeter during 5 to 8 days. Total power of the cell ( $P_{\text{Rext}}$ , W) was determined by

measuring cell voltage ( $V_{celda}$ , V) and external resistance ( $R_{ext}$ ,  $\Omega$ ), according to Equation (1), and current (I, A) is calculated from Ohm's law according to Equation (2) [12].

$$P_{R_{ext}} = V_{celda}^2 / R_{ext} \quad (1)$$

$$I = V_{celda} / R_{ext} \quad (2)$$

Output power is usually normalized with respect to anode surface area ( $S_A$ ,  $m^2$ ), because this is where the biological reaction occurs. The power density ( $DP_A$ ,  $W/m^2$ ) was calculated based on Equation (3) [17].

$$DP_A = V_{celda}^2 / (R_{ext} \times S_A) \quad (3)$$

### III. RESULTS

In this section, the SEM microphotographs and nitrogen adsorption-desorption isotherms at  $-196$  °C of the activated carbons are presented. Moreover, the activated carbons were used as electrodes in the anode chamber and their electrochemical performance in the MFC are presented.

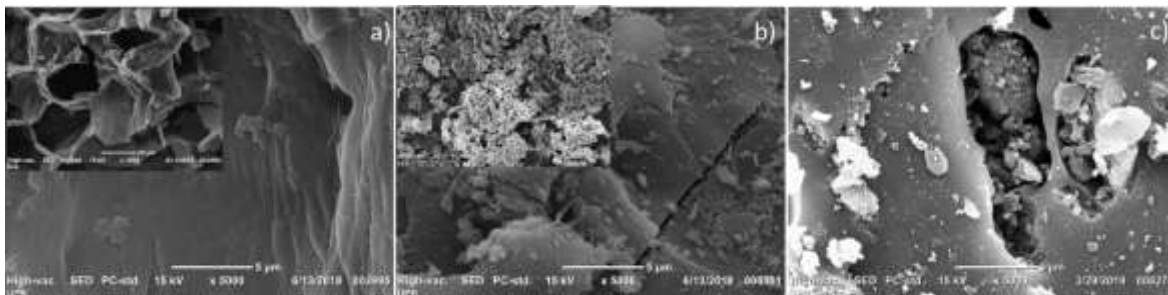
#### A. Anodic Chamber Electrodes

The morphology of the materials used as electrodes in the anode chamber was determined by SEM micrographs that are presented in Figure 2. Coffee husk surface is rough, irregular and undulating [18], and shows a plate morphology with a low specific surface area. The morphology of activated carbon from coffee cisco with  $H_3PO_4$  (Q) is observed in Figure 2a. The structure of sample Q is composed by “flakes” or dishes that form cavities and wide macropores, similar to the materials prepared by Maimulyanti et al. [19]. Its porous structure is made up of homogeneous pores (figure inserted in Figure 2) compared to the coffee husk activated with  $NaOH/CO_2$  by Ahmad et al. [18]. Also, sample Q shows deep pores compared with the coffee husk activated with  $ZnCl_2$  by Kosaiyakanon et al. [20], which surface has shallow and heterogeneous craters.

Figure 2b shows the morphology of activated carbon A, which appears to be a dense structure, with large, smooth, and cracked faces, with a slight presence of macropores (Figure 2b inserted). Finally, activated carbon P morphology (Figure

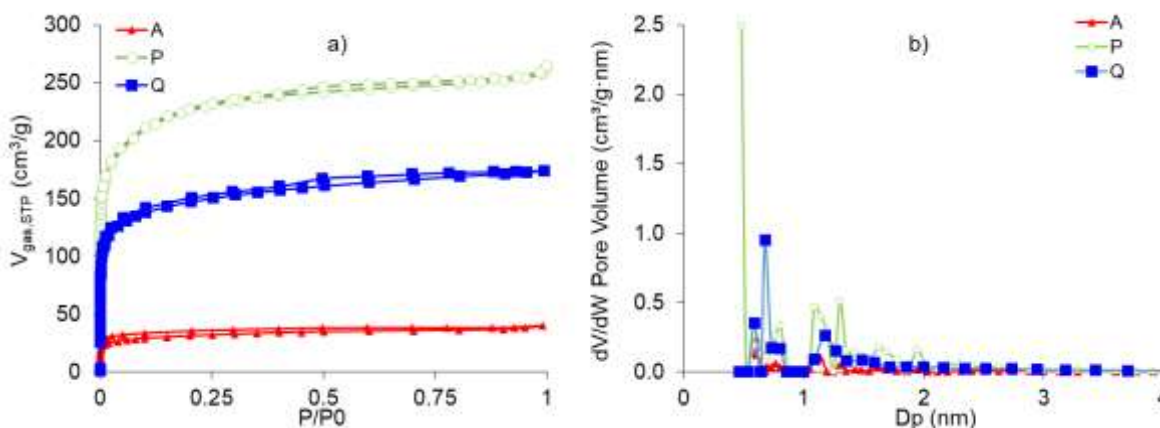


2c) shows large surfaces, with smooth sections and others with high roughness that promote the formation of a porous structure.



**Fig. 2.** SEM micrographs of the granular activated carbons a) Q and figure inserted at x1000, b) A and figure inserted at x800, and c) P.

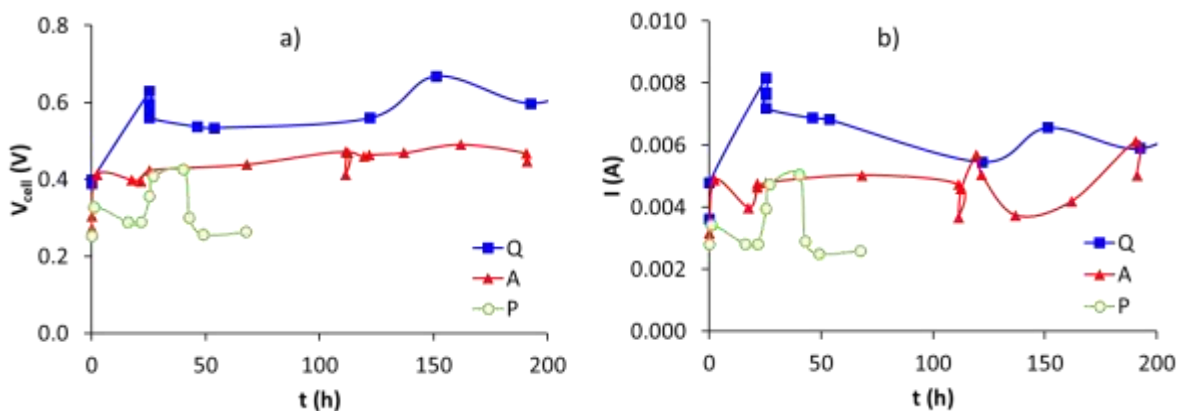
CAs presented mainly microporous characteristics, type I isotherms according to the IUPAC [21], with the presence of small mesopores in samples P and Q. Activated carbon P showed the highest value of  $S_{BET} = 816 \text{ m}^2 / \text{g}$ , followed by Q =  $541 \text{ m}^2/\text{g}$  and A =  $114 \text{ m}^2/\text{g}$ . Micropores size was very similar for the three activated carbons, varying between 1.20 and 1.31 nm, while sample Q had a greater external surface area of  $135 \text{ m}^2/\text{g}$  compared to samples P of  $62 \text{ m}^2/\text{g}$  and A of  $9.6 \text{ m}^2/\text{g}$ . Nitrogen adsorption-desorption isotherms at  $-196 \text{ }^\circ\text{C}$  are presented in Figure 3 and specific surface values and porous properties of the activated carbons are presented as supplementary material.



**Fig. 3.** a)  $\text{N}_2$  isotherms at  $-196 \text{ }^\circ\text{C}$  and b) the pore size distribution of granular activated carbons.

## B. Electrochemical Cell Characterization

Figure 4a shows the voltage values as a function of time. Sample Q showed the highest values followed by sample A and finally by sample P. These results may be related to a greater capillary force that allows migration of the aqueous medium towards the macropores and later mesopores that improve the wettability of the surface [11]. Voltage values and trend obtained were similar to those presented by Kim et al. [12]. In this work graphite cloth was used as an electrode. Furthermore, it is observed that the output voltage reaches a stable behaviour after 50 h. Figure 4b shows an increase in current, followed by a decrease as time goes on. This behaviour is similar to the results presented by Schoen [9], who used carbon fibers as electrodes in the anode chamber.

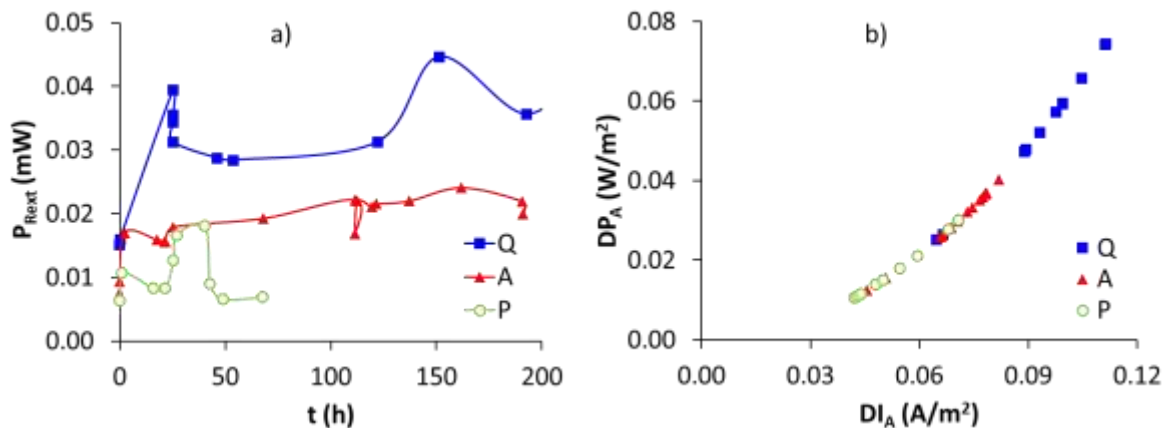


**Fig. 4.** a) Voltage and b) Cell current ( $I$ ) as a function of time of activated carbons granular at the anode.

## IV. DISCUSSION

It can be seen, in Figure 5a, that carbon Q showed the highest values of power of the anode. Two maximums were observed, one of 0.039 mW at 25 h and another of 0.045 at 150 h. When using carbon Q, the highest activity of the microorganism occurs during the first 30 h of the microbial fuel cell performance. Subsequently, a decrease in current was observed, which may be related to the diminishing of the substrate in the medium. As the microbial population decreases, the electron transport decreases too. Substrate availability has a significant influence on bacterial population growth and the formation of the biofilm in the anode, and

therefore in the performance of the power density and cell efficiency [6]. The increase in power after 120 h of cell operation may be related to the feeding of live microorganisms with the dead population.

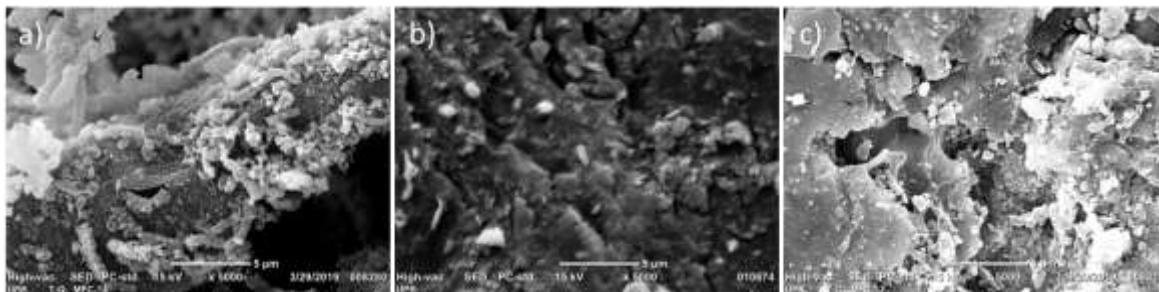


**Fig. 5.** a) Power versus time, and b) Power density versus cell current density for granular activated carbons at the anode.

Otherwise, P and A carbons presented power values between 0.008 mW and 0.018 mW, and between 0.006 mW and 0.024 mW, respectively. The increase and decrease in power presented by Q are also observed in the performance of coals P and A. Although the carbon P showed the highest specific surface area it showed the worst electrochemical performance, which indicates that not all the area provided by the micropores was in contact with the microorganism, the formation of micro-beds was less and therefore the transfer of electrons decreases. Some researchers use a lower external resistance (1  $k\Omega$ ) and larger external surface areas of the electrodes (25 - 50  $cm^2$ ), to maximize cell performance. The power density ( $DP_A$ ) respect to the anode current density ( $DI_A$ ) is directly proportional. Carbon Q showed the highest values, followed by A, and finally, by carbon P. These values were similar to Zhang et al. [22] and Cui et al. [11].

Biofilm formation change according to the kind of microorganism, its growth occurs in three stages: microbes, glycocalyx (secreted polymer matrix that protects bacteria on the surface) and a surface [23-24]. According to Yang [25], *S. putrefaciens* is one of the most versatile bacteria regarding its respiration. Furthermore, once the electrode is covered with biofilm, the donation of the

electrons could take place directly from the biofilm or it could act as a mediator between the electrode and the organic matter suspended in the medium.



**Fig. 6.** SEM microphotographs of activated carbons surface a) Q, b) A, and c) P after the electrochemical process.

It can be seen in Figure 6, those microorganism wastes remain on the surface of the three activated carbons, which can be associated with the biofilm. These wastes are not uniform and show a length between 1  $\mu\text{m}$  and 3  $\mu\text{m}$  (Measurements are presented in supplementary material). The *S. putrefaciens* microbeds are like to those presented by Cui et al. in composite materials from polyaniline, graphite cloth and carbon nanotubes [11]. Carbon Q (Figure 6a) showed a surface completely cover by rod-shaped microorganisms, and they have slightly adhered to the surface and some overlap on others, suggesting that this biofilm is made up of several communities of microbial cells [26]. The microorganisms adhered to surface of carbons A and P were spherical shape and in some parts of the surface were difficult to identify. The open structure or “flakes” morphology of the carbon Q allowed a greater biofilm formation and adherence to the surface, which improved the electrochemical performance of the cell.

## V. CONCLUSIONS

The anode material acts an important role in the production of electricity, since, depending on the nanostructure, the electron transfer and power density of the MFC can be improved. *S. putrefaciens* is an exoelectrogenic microorganism, that produces electricity that can use electrodes as electron acceptors. The macroporous morphology of activated carbon from coffee husk improved the

microbeds attachment on the surface of the electrode. This improves the metabolism of this microorganism through the MFC anode and promotes the extracellular transfer of electrons.

#### **AUTHOR'S CONTRIBUTION**

The main idea and work procedure were carried out by Zapata-Benabithé, Vanegas-Hernández, and Cardona-Aristizabal. Cardona-Aristizabal prepared the microbial culture, Zapata-Benabithé synthesized activated carbons Q and A. Configuration and electrochemical measurements of the microbial fuel cell were carried out by Vanegas-Hernández. The discussion was carried out by Zapata-Benabithé, and their checking by Vanegas-Hernández and Cardona-Aristizabal.

#### **FUNDING**

The project was financed by the Universidad Pontificia Bolivariana (UPB) through project No. 366B-03 / 15-19 from which these results were derived.

#### **ACKNOWLEDGMENTS**

The authors acknowledge to Laboratories of Materials from UPB for SEM analysis.

#### **REFERENCES**

- [1] T. González Estrada, and J. A. Valencia, Integración de las energías renovables no convencionales en Colombia. Bogotá D.C.: UPME, MINMINAS, BID, FMAM, 2015.
- [2] F. N. Jiménez-García, A. M. Restrepo-Franco, and L. F. Mulcúe-Nieto, "Estado de la investigación en energía en Colombia: una mirada desde los grupos de investigación," *Revista Facultad de Ingeniería*, vol. 28 (52), pp. 9-26, 2019. <https://doi.org/10.19053/01211129.v28.n52.2019.9651>
- [3] D. R. Lovley, "Bug juice: harvesting electricity with microorganisms.," *Nature Reviews Microbiology*, vol. 4 (7), pp. 497-508, Jul. 2006. <https://doi.org/10.1038/nrmicro1442>
- [4] P. Sengodan, and D. B. Hays, "Topic paper #13 Microbial Fuel Cells," *NPC Future Transportation Fuels Study: Advancing Technology for America's Transportation Future Study Topic Papers*. Texas, 2012.
- [5] I. Kim, K.-J. Chae, M.-J. Choi, and W. Verstraete, "Microbial Fuel Cells: Recent Advances, Bacterial Communities and Application Beyond Electricity Generation," *Environmental Engineering Research*, vol. 13 (2), pp. 51-65, 2008. <https://doi.org/10.4491/eer.2008.13.2.051>
- [6] D. Pant, G. Van Bogaert, L. Diels, and K. Vanbroekhoven, "A review of the substrates used in microbial fuel cells (MFCs) for sustainable energy production," *Bioresour. Technol.*, vol. 101 (6), pp. 1533-1543, Mar. 2010. <https://doi.org/10.1016/j.biortech.2009.10.017>

- [7] M. Osman, A. Shah, and F. Walsh, "Recent progress and continuing challenges in bio-fuel cells. Part II: Microbial," *Biosensors and Bioelectronics*, vol. 26 (3), pp. 953-963, Nov. 2010. <https://doi.org/10.1016/j.bios.2010.08.057>
- [8] A. Falcón, J. E. Lozano, and K. Juárez, "Bioelectricidad," *Rev. la Soc. Mex. Biotecnol. y Bioingeniería A.C.*, vol. 13 (3), pp. 62-78, 2009.
- [9] A. R. Schoen, "Carbon fiber electrode as an electron acceptor for a microbial fuel cell using geobacter," *Cantaurus J. McPherson Coll. Sci.*, vol. 15, pp. 24-26, 2007.
- [10] K. Scott, G. Rimbu, K. Katuri, K. Prasad, and I. Head, "Application of modified carbon anodes in microbial fuel cells," *Process Safety and Environmental Protection*, vol. 85 (5), pp. 481-488, 2006. <https://doi.org/10.1205/psep07018>
- [11] H. F. Cui, L. Du, P. B. Guo, B. Zhu, and J. H. T. Luong, "Controlled modification of carbon nanotubes and polyaniline on macroporous graphite felt for high-performance microbial fuel cell anode," *Journal of Power Sources*, vol. 283, pp. 46-53, 2015. <https://doi.org/10.1016/j.jpowsour.2015.02.088>
- [12] H. J. Kim, H. S. Park, M. S. Hyun, I. S. Chang, M. Kim, and B. H. Kim, "A mediator-less microbial fuel cell using a metal reducing bacterium, *Shewanella putrefaciens*," *Enzyme and Microbial Technology*, vol. 30 (2), pp. 145-152, 2002. [https://doi.org/10.1016/S0141-0229\(01\)00478-1](https://doi.org/10.1016/S0141-0229(01)00478-1)
- [13] C. K. Lee, A. J. Kim, G. S. Santos, P. Y. Lai, S. Y. Lee, D. F. Qiao, J. De Anda, T. D. Young, Y. Chen, A. R. Rowe, K. H. Neelson, P. S. Weiss, and G. C. L. Wong, "Evolution of Cell Size Homeostasis and Growth Rate Diversity during Initial Surface Colonization of *Shewanella oneidensis*," *ACS Nano*, vol. 10 (10), pp. 9183-9192, 2016. <https://doi.org/10.1021/acs.nano.6b05123>
- [14] E. Marsili, D. B. Baron, I. D. Shikhare, D. Coursolle, J. A. Gralnick, and D. R. Bond, "Shewanella secretes flavins that mediate extracellular electron transfer," *PNAS*, vol. 105 (10), pp. 3968-3973, 2008. <https://doi.org/10.1073/pnas.0710525105>
- [15] H. H. Hau, and J. A. Gralnick, "Ecology and Biotechnology of the Genus *Shewanella*," *Annual Review of Microbiology*, vol. 61 (1), pp. 237-258, 2007. <https://doi.org/10.1146/annurev.micro.61.080706.093257>
- [16] S. You, Q. Zhao, J. Zhang, J. Jiang, and S. Zhao, "A microbial fuel cell using permanganate as the cathodic electron acceptor," *Journal of Power Sources*, vol. 162 (2), pp. 1409-1415, 2006. <https://doi.org/10.1016/j.jpowsour.2006.07.063>
- [17] B. E. Logan, B. Hamelers, R. Rozendal, U. Schröder, J. Keller, S. Freguia, P. Aelterman, W. Verstraete, and K. Rabaey, "Microbial Fuel Cells: Methodology and Technology," *Environmental Science and Technology*, vol. 40 (17), pp. 5181-5192, 2006. <https://doi.org/10.1021/es0605016>
- [18] M. A. Ahmad, and N. K. Rahman, "Equilibrium, kinetics and thermodynamic of Remazol Brilliant Orange 3R dye adsorption on coffee husk-based activated carbon," *Chemical Engineering Journal*, vol. 170 (1), pp. 154-161, 2011. <https://doi.org/10.1016/j.cej.2011.03.045>
- [19] A. Maimulyanti, A. R. Prihadi, T. Rosita, and I. Safrudin, "Adsorption and recovery of aroma compounds from wastewater of clove oil distillation using coffee husk biosorbent," *ScienceAsia*, vol. 45 (5), pp. 446-451, 2019. <https://doi.org/10.2306/scienceasia1513-1874.2019.45.446>
- [20] L. C. A. Oliveira, E. Pereira, I. R. Guimaraes, A. Vallone, M. Pereira, J. P. Mesquita, and K. Sapag, "Preparation of activated carbons from coffee husks utilizing FeCl<sub>3</sub> and ZnCl<sub>2</sub> as activating agents," *Journal of Hazardous Materials*, vol. 165 (1-3), pp. 87-94, Jun. 2009. <https://doi.org/10.1016/j.jhazmat.2008.09.064>

- [21] M. Thommes, K. Kaneko, A. V. Neimark, J. P. Olivier, F. Rodriguez-Reinoso, J. Rouquerol, and K. S. W. Sing, "Physisorption of gases, with special reference to the evaluation of surface area and pore size distribution (IUPAC Technical Report)," *Pure and Applied Chemistry*, vol. 87 (9-10), pp. 1051-1069, 2015. <https://doi.org/10.1515/pac-2014-1117>
- [22] X. Zhang, J. Shi, P. Liang, J. Wei, X. Huang, C. Zhang, and B. E. Logan, "Power generation by packed-bed air-cathode microbial fuel cells," *Bioresource Technology*, vol. 142, pp. 109-114, Aug. 2013. <https://doi.org/10.1016/j.biortech.2013.05.014>
- [23] W. Michael Dunne Jr., "Bacterial Adhesion: Seen Any Good Biofilms Lately?," *Clinical Microbiology Reviews*, vol. 15 (2), pp. 155-166, 2002. <https://doi.org/10.1128/cmr.15.2.155-166.2002>
- [24] J. C. Biffinger, J. Pietron, R. Ray, B. Little, and B. R. Ringeisen, "A biofilm enhanced miniature microbial fuel cell using *Shewanella oneidensis* DSP10 and oxygen reduction cathodes," *Biosensors and Bioelectronics*, vol. 22 (8), pp. 1672-1679, 2007. <https://doi.org/10.1016/j.bios.2006.07.027>
- [25] Y. Yang, G. Sun, J. Guo, and M. Xu, "Differential biofilms characteristics of *Shewanella* decolorationis microbial fuel cells under open and closed-circuit conditions," *Bioresource Technology*, vol. 102 (14), pp. 7093-7098, 2011. <https://doi.org/10.1016/j.biortech.2011.04.073>
- [26] T. H. Pham, P. Aelterman, and W. Verstraete, "Bioanode performance in bioelectrochemical systems: recent improvements and prospects," *Trends in Biotechnology*, vol. 27 (3), pp. 168-178, Mar. 2009. <https://doi.org/10.1016/j.tibtech.2008.11.005>